REVIEW

Open Access



Exploring the potential of nanomaterials (NMs) as diagnostic tools and disease resistance for crop pathogens

Muhammad Jabran¹, Muhammad Amjad Ali³, Saima Muzammil⁴, Adil Zahoor⁵, Faizan Ali³, Sarfaraz Hussain⁶, Ghulam Muhae-Ud-Din¹, Munazza liaz⁷ and Li Gao^{1,2*}

Abstract

Food crops are attacked by microbial pathogens and insect pests, leading to significant yield reductions and economic losses. Conventional disease diagnosis and management approaches often fail to provide rapid and ecofriendly solutions. In the current situation, nanomaterials (NMs) serve a valuable role in both managing emerging pathogens and monitoring overall plant health. Nanotechnology has transformed the biotechnology industry including agriculture with specific applications such as nano-fungicides, nano-bactericides, and nano-pesticides. This review focuses on the use of various nanomaterials, including inorganic materials such as Ag, ZnO, CuO, and CeO, as well as carbon-based nanoparticles, nanotubes, nanowires, and nano-capsules. The application of NMs holds the potential to address various challenges in food security through novel applications like advanced nano-biosensors for rapid pathogen detection and targeted disease management strategies. This includes the potential to minimize reliance on chemical inputs and contribute to more sustainable agricultural practices. Nanomaterials (NMs) promise to deliver plant hormones and signaling molecules to plants, enhancing resistance inducers against major crop pathogens. NMs against newly arising pathogens through reactive oxygen generation, membrane damage, and biochemical interference are also reviewed. However, challenges regarding the stability, toxicity, and environmental impacts of NMs are discussed, along with recommendations on green synthesis and functionalization approaches. This article aims to investigate the role of nanomaterials (NMs) in managing emerging pathogens and monitoring overall crop health offering an insightful outlook for future generations. Further biosafety aspects and larger-scale validation of NM-based applications could enable their commercialization for improving global food security.

Keywords Nanomaterials, Disease resistance, Plant defense system, Plant pathogens, Disease identification, Ecofriendly

*Correspondence: Li Gao xiaogaosx@hotmail.com; lgao@ippcaas.cn Full list of author information is available at the end of the article



© The Author(s) 2024. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativeco mmons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.



Introduction

The global demand for food crops is approximately 2-3billion tons, which is needed to feed the world's 7.9 billion people. However, this demand is expected to increase by 50% in the coming years, creating a significant challenge for agricultural production to keep up [1]. Food crops are facing severe threats from plant pathogens and the impacts of climate change [2]. Plant diseases are major barriers to agricultural development and food security, lowering crop yields, as well as the quality of the production in storage [1]. For instance, a historic epidemic of Phytophthora infestans that caused potato late blight disease in Ireland had substantial cultural and economic repercussions, leading to population movement and starvation in the 1840s [3]. Pathogens are responsible for significant agricultural output losses around 20%-40% [4], prompting ongoing research into managing fungal, viral, nematode, and bacterial diseases. Plant pathogens have evolved mechanisms to evade plant defense systems by secreting effector proteins that interact with specific plant target proteins, thereby suppressing plant immunity and facilitating disease development [5]. Plant diseases impart a substantial yield penalty on major global food crops. A recent study estimated average yield loss from diseases across potato (17.2%), soybean (21.4%), wheat (21.5%), maize (22.5%), and rice (30.0%) comprising over half of global calorie consumption crops. Worryingly, the maximum losses happened in food-insecure areas with high populations that also face emerging or re-emerging diseases aggravated by climate change. Targeted efforts to encourage sustainable agricultural productivity through minimizing yield gaps from pathogens will be critical to global food security [1]. In the past, agrochemicals have mitigated plant diseases, but their overuse has led to resistance, environmental harm, and unintended species accumulation. To address these challenges, alternative disease control strategies are imperative due to their profound implications for human health and the environment [6]. Novel control measures such as nanotechnology include nanomaterials, nanoparticles, nanocarbon materials, agrochemical-based nanomaterials [7], nano-biosensors, and small detection devices that could be used for the detection and management of plant pathogens. NMs can significantly contribute to revolutionizing agrotechnology because they are more efficient, defendable, and durable than currently

used synthetic chemical products. For example, the development of nanosensors for early disease detection [8], nano-fertilizers, nanopesticides, nanoherbicides [9], growth promoters, and antimicrobials presents a hopeful path for sustainable crop protection and productivity improvement. Collectively, NMs present versatile opportunities to transform plant disease management and agriculture worldwide. These materials enable site-specific and controlled nutrient release, improving plant uptake efficacy while producing disease resistance and mitigating environmental impacts [10]. Various nanostructured materials, including nanoclays, carbon nanodots, nanotubes, nanofibers, carbon-based nanomaterials, and polymeric nanoparticles, serve as promising carriers for nutrients [11]. In recent years, the use of nanomaterials has been considered a suitable solution to control plant pests, including insects, fungi, and weeds [12]. Furthermore, NMs can act as stimulators for plant defense mechanisms helping to prevent disease [13]. For example, engineered nanomaterials (ENMs) were applied to a 5-mL suspension containing 50 or 200 mg L^{-1} concentrations of two metal-based nanoparticles (Fe_2O_3 or TiO_2) and two carbon-based nanomaterials (MWCNTs or C60). These ENMs were sprayed onto tobacco leaves every day for 21 days and fully developed young leaves were inoculated with turnip mosaic virus TuMV tagged with green fluorescent protein (GFP). The fluorescence images of TuMV abundance on the leaf surfaces indicated that NMs exhibited a substantial inhibitory effect on viral proliferation, as evidenced by reduced fluorescence intensity on the newly emerged leaves. Moreover, a noteworthy reduction ranging from 15 to 60% was observed in the relative quantity of TuMV coat proteins, providing additional insights into the mechanisms through which NMs exert their suppressive influence on viral infection [14]. In recent studies, nanoparticles, including zinc oxide (ZnO), titanium dioxide (TiO₂), gold (Au), silver (Ag), cerium oxide (CeO_2), silica oxide (SNP), and copper (Cu), have found extensive application in crop fields [15]. In prior studies, it was observed that metal oxide nanomaterials such as TiO₂ NPs and CuO NPs, as well as carbon nanomaterials like reduced graphene oxide (GO) and multi-wall carbon nanotubes (MWCNTs), exhibited distinct inhibition of the fungus Podosphaera pannosa when applied at high concentrations 200 mg/L in preventing infestation of rose leaves. However, when used at lower concentrations 50 mg/L, only CuO NPs demonstrated inhibitory effects on P. pannosa. Hence, a multidisciplinary technique is required to examine the impact of nanomaterials on plant diseases, including gene expression, transcriptomic analysis, proteomics, and secondary metabolites [16]. Furthermore, NMs have the capability to influence plant biochemical

and molecular responses, such as activating antioxidant defense systems, accumulating osmolytes and hormones, and regulating gene expression. These mechanisms collectively enhance plant resilience against biotic stresses [17]. For example, nanoparticles like TiO2 induce the expression of P5CS1 gene, enhancing proline synthesis and stress resilience in plants [18-20]. Moreover, NPs (ZnO, Se, Si) boost antioxidant enzyme activity such as APX, CAT, and SOD, reducing drought-induced oxidative damage [21]. NMs have demonstrated potential antimicrobial effects through the modulation of several intracellular pathways. Their antimicrobial mode of action may involve suppression of ATP synthesis, disruption of cell membranes, alterations in membrane permeability, inhibition of enzyme production, interference with electron transport processes, disruption of cytochrome interactions, and enhancement of reactive oxygen species generation. Collectively, these diverse cellular impacts of NMs can effectively control microbial growth and viability through both membrane and metabolic perturbations [22]. The contribution of nanomaterials and nanotechnology in the agricultural field is still in its infancy. Therefore, a system-level approach is required to provide more precise information on the role of nanoparticles against plant pathogens. The use of several nanomaterials in crop protection for disease resistance and early detection via triggering defense responses to pathogens is reviewed in this article. Moreover, plant disease monitoring using nano-biosensors is an innovative and highly promising approach for the early detection and management of plant diseases. Nano-biosensors operate on various principles, including surface plasmon resonance, fluorescence, electrochemistry, and fieldeffect transistor technology. These sensors employ biorecognition elements such as antibodies, and DNA probes to selectively bind to pathogen-specific molecules ensuring high sensitivity and specificity. They can detect various plant pathogens from bacteria to viruses and fungi, offering advantages such as high sensitivity, specificity, real-time monitoring, and cost-effectiveness. For instance, case studies highlight their application in detecting plant pathogenic bacteria such as Xanthomonas and Ralstonia spp. [23] and fungal pathogens such as Fusarium and Botrytis spp. [24]. Despite their potential, challenges such as affordability and integration into agricultural practices remain to be addressed. Nevertheless, ongoing research improves biorecognition elements and sensor platforms making nano-biosensors a valuable tool for early and accurate plant disease monitoring. This technology empowers farmers and agriculturalists to proactively manage crop health, ultimately reducing yield losses. Furthermore, this review summarizes the effectiveness of different NMs used against various plant

diseases and the safe applications of NMs in food crops. The review also elucidates the various biochemical and molecular mechanisms facilitated by nanoparticles (NPs) that promote pathogen resistance in plants. While existing research offers concise insight into NP characteristics, synthesis methods, and their contribution to pathogen detection and prevention, gaps remain in understanding their potential drawbacks in pathogenesis [25]. Therefore, this review aims to address existing knowledge gaps and highlight promising future research directions concerning the use of specific NMs in plant disease diagnosis and disease resistance. Areas of focus include exploration of biosafety considerations associated with NM applications and potential detrimental impacts on crop health. Understanding biosafety profiles and adverse effects is crucial to enabling safe development and deployment of NM technologies in agriculture. The review also discusses optimization of NM properties for effective disease detection and management while minimizing environmental risks. Overall, the review seeks to provide insights and perspectives to guide continued advancement of safe and sustainable NM solutions for improved plant health and productivity.

Nanomaterial synthesis and characterization for crop plants

Nanotechnology produces nanoscale materials called nanoparticles or nanomaterials in various forms, such as ferromagnetic, ferroelectric, and superconducting materials. The characteristics of nanomaterials are dependent on their size, charge, and shape. Nanomaterials in crop plants have a size typically between 1 and 100 nm and can be synthesized using different methods depending on the material and application. They can be classified into inorganic, carbon, organic, and composite nanomaterials. Nanoparticles can be produced from metal or metal oxide using physical, chemical, and biological procedures, with the biological methods being environmentally safe. In the realm of nanomaterial synthesis, various methods including chemical reduction, sol-gel synthesis, and green synthesis, are employed for nanomaterial synthesis. Among them, green synthesis utilizing biological entities like plants and fungi to reduce metal ions is remarkable for its sustainability. Recent studies have successfully synthesized gold nanoparticles (Au-NPs) through this eco-friendly method [26]. Green synthesis of nanoparticles, which uses plant extracts and microorganisms, is frequently used to produce metal-based nanoparticles [27]. Numerous plants have been utilized for green synthesis of nanomaterials [28]. For example, a study evaluated 109 plant species from Middle Eastern traditional medicine, analyzing dried samples of different plant parts including bark, bulb, flower, fruit, gum, leaf, petiole, rhizome, root, seed, stamen, and above-ground parts. Out of 117 plant parts from 109 species across 54 families, 102 extracts showed the bio reduction of Au^{3+} to Au^{0} . This study unveiled 37 new plant species capable of AuNP synthesis [29]. According to the literature, leaf extracts from numerous plants including Cocos nucifera [30], Psidium guajava, Garcinia mangostana [31], Ocimum sanctum, Bryophyllum spp., Cyperus spp., Hydrilla spp., Rosa rugosa, and Chenopodium album [32], Azadirachta indica A. Juss [33], Eucalyptus globules [34, 35], etc., have been used to produce silver, gold, zinc, and copper, iron nanoparticles. Nanoparticles produced by plants are more stable and their synthesis rate is faster than that of microorganisms. Plant-based nanoparticles have been proven to be more beneficial due to the presence of biomolecule variability [36]. The plant biomolecules such as alkaloids, phenolics, terpenoids and proteins that leach out during nanoparticle synthesis play a critical role in modulating the NP size, shape, dispersity, and surface properties [37]. They form a protective biomolecular layer or "halo" around the NPs which likely facilitates their interactions with target pathogens [38]. Plant-based materials play a crucial role in green nanomaterial synthesis, using phytochemicals as reducing and capping agents, offering an ecofriendly alternative with enhanced biological effects and interactions with living systems. After the nanoparticles have been synthesized, they must be purified and characterized before being used to confirm their morpho-physical properties that may enhance their efficacy. Various techniques are used for this purpose, each tailored to address specific aspects of purification. Centrifugation, for instance, capitalizes on size and density inequalities to effectively separate nanomaterials from aggregates. Filtration with specific pore-sized membranes eliminates larger contaminants. On the other hand, dialysis aids facilitate the in the removal of small molecules and ions through selective diffusion across membranes. Precipitation methods, including solvent and co-precipitation separate nanomaterials from excess reagents. Additionally, surface modification strategies involving the use of ligands or coatings prevent aggregation and boost colloidal stability. The choice of purification method is influenced by nanomaterial properties, intended use, and desired purity levels that is guided by quality control measures [39]. Several approaches are involved in characterizing the size, shape, crystal structure, and atomic composition. The size and shape of nanomaterials are defined using innovative microscopy methods, including scanning electron microscopy (SEM), UV-visible spectroscopy [40], transmission electron microscopy (TEM), atomic force

microscopy (AFM), X-ray fluorescence microscopy (XFM), and scanning transmission X-ray microscopy (STXM) [41]. The different structural characteristics of nanoparticles were determined using nuclear magnetic resonance (NMR). Another essential approach for nanoparticle characterization is X-ray diffraction (XRD). Likewise, the crystalline structures of nanoparticles in various nanomaterials can be revealed using XRD. Furthermore, the charge of NPs is crucial. For example, nanoparticles can carry either a positive (cationic) or negative (anionic) charge, as determined by the type of molecules on their surfaces. This surface charge can be deliberately adjusted during nanoparticle synthesis or functionalization processes [42]. Nanomaterials are rapidly acquiring interest from several scientific segments; therefore, there is a need for the development of novel, cost-effective, and easy approaches for the largescale synthesis of nanomaterials.

Applications of nanomaterials in agriculture

Nanomaterials possess diverse physicochemical characteristics due to their small size, exhibiting higher reactivity, biochemical activity, and solubility owing to their elevated surface-to-volume ratio. Inorganic nanoparticles, particularly metal oxides like ZnO, TiO2, CuO, and CeO2, are predominantly employed in studies (79%), with metal nanoparticles, notably Ag, representing (25%). Carbon-based nanoparticles are less utilized, accounting for only 10% of studies [43]. These examples highlight the diverse real-world applications of nanomaterials in various aspects of agricultural management, including nutrient delivery, pest and disease control, diagnostics, and plant growth enhancement. Further, NMs have been explored for their potential in enhancing crop disease resistance by modulating plant biochemical and molecular responses, triggering the upregulation of defense-related genes. Several applications of NMs have been explored for controlled release of agrochemicals and nutrients, particularly micronutrients such as Fe, Mn, Zn, Cu, K, Ca, P, etc., enhancing plant biomass and growth [44, 45]. Mg nanoparticles (Mg-NFs) not only enhance seed germination and seedling growth attributes, but also serve as activators. Recent studies have highlighted the beneficial effects of various NPs, including carbon nanotubes (CNTs), silicon dioxide (SiO2), zinc oxide (ZnO), titanium dioxide (TiO2), and gold (Au) NPs, in promoting seed germination in wheat, pearl millet, tomato, soybean, barley, rice, and maize. For example, a study investigated the impacts of nano-biofertilizer on tomato crops affected by Ralstonia solanacearum caused bacterial wilt disease and its pest-resistant function against wilt disease [46].

Nanomaterials for enhancing crop resistance to pathogens

Approximately 14% of crops in the world are damaged by infectious diseases caused by plant pathogens, and yield losses could be as high as 20-40% globally [47]. Major cereal crops in the world, such as wheat, rice, barley, and maize, can be easily infected by fungal diseases. Fungal pathogens pose a significant threat to global crop production, accounting 70% of plant diseases. In addition, fungal diseases caused a significant loss of more than 3.41 million tons of wheat in China between 2000 and 2018 [48]. Nanomaterials are an integrated and sustainable approach because of their small size, nanoparticles (NPs) easily penetrate plant pathogens and can affect their disease-causing ability. More than 90% of applied pesticides are either lost in the environment or miss the target regions/microbes for effective disease control. This not only causes harmful impacts on the environment, but also increases the overall production cost for farmers. Therefore, one difficult area of agricultural research that still must be applied is the development of innovative crop protection formulations [10]. Nanomaterials have revolutionized agriculture by improving plant disease resistance. Some examples of how nanomaterials are being used to enhance plant resistance to pathogens have been reported [49]. A recent study investigates the efficacy of five distinct nanoparticles (NPs), namely Co_3O_4 CuO, Fe₃O₄, NiO, and ZnO, in combating Fusarium wilt and promoting common bean plant growth. In vivo experiments demonstrated that all NPs significantly improved resistance with respective disease control values of 92.84% (therapeutic) and 82.77% (protective). The plants were grown under greenhouse conditions. These results underscore the potential of nanomaterials in agriculture as nano-fungicides and nano-fertilizers with promising implications for sustainable agriculture and environmental preservation [50]. For instance, CuO nanoparticles have been used to improve disease resistance against Fusarium crown and root rot, Fusarium wilt, and Verticillium wilt in various plants. Copper and silver NP compounds are highly effective in eliminating several fungal pathogens, including Aspergillus carbonarius, Aspergillus fumigatus, Aspergillus niger, Aspergillus oryzae, Candida albicans and Cryptococcus neoformans [51]. Similarly, the antifungal properties of Ag NPs and CuO have been reported to suppress the growth of powdery mildew in different crops. Similarly, the effect of nanoparticles has been studied on Ustilaginoidea virens caused by false smut infection of rice [52]. Recent attention has focused on eco-friendly nanoparticle

production. This study successfully synthesized cerium oxide nanoparticles (CeO₂ NPs) using a green method involving quinoa leaf extract. The NPs were characterized as spherical clusters with sizes ranging from 7 to 10 nm. Testing of two wheat varieties revealed that higher CeO₂ NPs concentrations significantly reduced disease severity and incidence, particularly at 100 mg/L. These findings suggest promising antifungal potential for CeO₂ NPs against Ustilago tritici, offering a potential solution for global crop protection [53]. However, there is still a need for a critical review to discuss the most recent developments in this field and to clarify research areas on their ecological safety and difficult gaps. Overall, the application of nanomaterials against plant pathogens is a promising area of research with the potential to revolutionize the way we protect crops and increase yields. The different NMs could be used via foliar, seed treatment, soil application, and plant root applications against the plant pathogens discussed in Fig. 1.

Copper nanoparticles (Cu NPs)

Copper-based nanoparticles have garnered significant attention for their potential in controlling a wide range of plant pathogens, including bacteria and fungi. Notably, several studies have demonstrated that copper nanoparticles can serve as a more effective and achieving comparable efficacy at lower concentrations. In addition, copper nanoparticles can be applied in a variety of forms, including as a foliar spray, seed coating, or soil amendment, making them versatile for different cropping systems and plant growth stages [54]. Researchers have been particularly fascinated by Cu NPs because of their distinctive biological, chemical, physical, and antibacterial properties. In addition, iron- and copper-based nanoparticles react with peroxides in the environment, generating free radicals that are highly toxic to microorganisms [55]. For instance, studies have demonstrated the significant antifungal activity of Cu NPs against common crop pathogens such as F. oxysporum, F. culmorum, and F. equiseti,



Fig. 1 Illustration describes the application of nanomaterials (NMs). The schematic represents the different ways in which nanomaterials can be applied to protect plants from pathogens. The most common method is through foliar spray, where nanoparticles penetrate plant tissue and provide long-lasting protection against fungal infections and insect pests. Another application is through seed treatments, where nanoparticles can be used to protect seeds from soil-borne pathogens before they are planted. These nanoparticles can form a physical barrier around the seed, preventing pathogens from penetrating and infecting the seed. By binding to the pathogen and preventing it from infecting the plant roots, these nanoparticles can help protect the plant from disease

[56]. Further, the study presents a promising approach for the development of environmentally friendly copperbased fungicides using neem leaf extract. It effectively controlled the pathogens of apple orchards, including *Alternaria mali, Diplodia seriata*, and *Botryosphaeria dothidea* [34, 35]. However, it is important to note that the use of copper nanoparticles for plant pathogen control is still in its early stages, and more research is needed to fully understand their effect on plants and the environment. Also, overuse of copper nanoparticles can lead to copper toxicity, which is harmful to plants.

Silver nanoparticles (Ag NPs)

The most prevalent inorganic NPs used for antimicrobial properties are Ag NPs. These were the first nanoparticles to be used in agriculture to combat plant pathogens [57]. Silver is known for its broad-spectrum antimicrobial properties and its ability to disrupt the cell membrane of pathogens. At the molecular level, research has shown that Ag NPs can enhance plant disease resistance by inducing the production of important defense compounds in plants. Studies report Ag NP treatment leads to increased biosynthesis of phytoalexins, phenolic compounds, terpenoids and polyphenols known to play critical roles in the plant defense response against pathogens [58]. Various soil-borne diseases have been controlled by using silver nanoparticles including Phytophthora parasitica, Meloidogyne spp., and Fusarium spp. [59]. During field tests, a reduction in the symptoms of powdery mildew in cucumber and pumpkin caused by the fungi Golovinomyces cichoracearum and Sphaerotheca fusca was reported after Ag NP sprays at 10-100 mg/L [60]. Additionally, at a concentration of 15 mg/mL, the biosynthesized Ag-NPs showed outstanding inhibitory effectiveness against Curvularia lunata, Rhizoctonia solani, Macrophomina phaseolina, Sclerotinia sclerotiorum, Alternaria alternata, and Botrytis cinerea [61]. Furthermore, biosynthesized nanoparticles from various sources have shown considerable antibacterial activity against pathogenic bacteria in vitro and in vivo. Similarly, plant pathogenic fungi such as Bipolaris sorokiniana and Mag*naporthe grisea* have been reported to be suppressed by several types of silver ions and nanoparticles [62]. However, silver nanoparticles (AgNPs) were synthesized using aqueous leaf extract obtained from Aloysia citrodora and evaluated for their antifungal activity against soil-borne and airborne pathogens, including Pythium aphanidermatum, Paecillomyces formosus, Macrophomina phaseolina, and Botrytis cinerea. The study demonstrated significant inhibition of mycelial growth. These findings suggest that environmentally friendly biosynthesized Ag-NPs possess more potential to combat phytopathogens in the agricultural sector [7, 63].

Zinc oxide nanoparticles (ZnO NPs)

ZnO-NPs as a sustainable alternative for controlling harmful plant pathogens and safeguarding global food security. In the literature, they have been found to be effective against fungal pathogens such as Fusarium oxysporum and Botrytis cinerea, bacterial pathogens such as Pseudomonas syringae, and viral pathogens such as tobacco mosaic virus (TMV) [64]. At the molecular level, ZnO NPs can interact with various cellular components in plant cells to enhance disease resistance. These interactions include interactions with plant cell membranes, proteins, DNA, and other cellular components that can inhibit the growth and reproduction of disease-causing microorganisms. Mechanically, ZnO NPs can also induce the production of reactive oxygen species (ROS) within plant cells, which can inhibit the growth of pathogens. Numerous scientists have studied zinc oxide nanoparticles and discovered that they effectively reduce fungal growth in crops. For example, ZnO NPs produced distortion in fungal hyphae and inhibited the production of conidiophores and conidia, according to scanning electron microscopy (SEM) photographs and Raman spectra [65]. Similarly, the effectiveness of zinc compounds in preventing wheat deoxynivalenol production and Fusarium head blight was investigated. The impact of presowing seed application with metal nanoparticles (Zn, Ag, Fe, Mn, and Cu) on the development of resistance in wheat seedlings infected with Pseudocercosporella herpotrichoides was demonstrated by [66]. In a recent study, ZnO NPs were employed to protect tomato plants against Fusarium wilt. These ZnO NPs exhibited significant potential as inducers of plant physiological immunity against Fusarium wilt, reducing disease incidence by 28.57% and providing high protection by 67.99% against F. oxysporum. Furthermore, they enhanced various growth parameters and biochemical compounds indicating their effectiveness in controlling and fortifying plants against fusarial infection [67]. Hence, the exact molecular and mechanistic aspects of zinc oxide nanoparticles in plant disease resistance are still not fully understood, and more research is needed to fully understand their mechanisms of action.

Chitosan nanoparticles

Chitosan nanoparticles show promise as biopesticides for crop protection due to their broad-spectrum antifungal activity, biocompatibility with plant materials, and low environmental toxicity. Their reported efficacy against an array of phytopathogenic fungi combined with benign safety profile make chitosan nanoparticles an attractive option for sustainable disease control in agricultural systems [68]. Due to its sustainable and safe properties, chitosan has become the material of

choice to produce nanoparticles in agricultural fields. Previous studies suggest their high molecular weight, cationic charge, and surface hydrophobicity enable interaction with negatively charged fungal cell membranes. This interaction is pondered to disrupt membrane integrity through mechanisms such as increased permeability. For example, in a study by Zheng et al. [69], chitosan effectively enhanced resistance to *Phy*tophthora infestans in potted potatoes. In vitro studies have shown that chitosan treatment can significantly reduce leaf lesion sizes. Chitosan nanoparticles exhibited substantially smaller lesion sizes compared to untreated control leaves when assessed 5-7 days postinfection. Chitosan at 0.5 g/L provided 46.0% protection, which was slightly higher than 0.25 g/L (35.5%). Furthermore, transcriptomics revealed chitosan-producing resistance, as confirmed by qRT-PCR analysis. A total of 11,410 differentially expressed genes (DEGs) were identified, with 6026 genes showing upregulation and 5384 genes showing downregulation. Chitosan appeared to upregulate these DEGs, indicating a positive response of potatoes to chitosan treatment. It also induced ROS- and SA-related gene expression, confirming disease resistance against P. infestans. Chitosan is essential to induce resistance in different crops. Studies have demonstrated that chitosan nanoparticles have antimicrobial activities and reduced disease severity against many plant pathogens, including Fusarium graminearum, F. oxysporum, Phytophthora infestans, Xanthomonas campestris, Erwinia carotovora, Pseudomonas syringae, and Clavibacter michiganensis [70]. For example, chitosan NPs have been found to be resistant against fungal pathogens including Aspergillus niger, Alternaria alternata, Rhizopus oryzae, Phomopsis asparagi, and Rhizopus stolonifer [71, 72]. In the literature, studies have shown that chitosan NPs can inhibit the growth of the plant pathogen Clavibacter michiganensis by disrupting its cell wall. Chitosan NPs can increase the production of defense-related enzymes such as peroxidases and catalases leading to enhanced resistance to pathogens [73]. Further, chitosan NPs have been found to induce the production of ROS and systemic acquired resistance (SAR) [74]. Similarly, Cu-chitosan nanomaterials have been tested for their ability to promote plant growth and enhance systemic resistance to the Curvularia leaf spot (CLS) disease of maize. Higher antioxidant (superoxide dismutase and peroxidase) and enzyme activities were evidence of a significant response in plants exposed to Cu-chitosan NPs [75]. However, it is important to keep in mind that the understanding of how chitosan nanoparticles work at the molecular and mechanistic level to enhance plant disease resistance is still ongoing, and further research is needed to fully understand their impact on plants and the environment. Several nanomaterials used for disease resistance in different crops are summarized in Table 1.

Nanomaterials as activators of immune responses in crops against pathogens

Plants possess natural immune systems that allow them to detect invading pathogens and launch tailored defense responses. Using pattern recognition receptors, plants can recognize signatures associated with bacteria and fungi. Upon pathogen perception, complex signaling networks are activated that initiate multifaceted immune responses aimed at protecting the plant and limiting infection spread [106]. They activate defense mechanisms including chemical responses and gene regulation which all work together to protect plants from diseases and infections. This process leads to the production of reactive oxygen species (ROS), antioxidants, and stimulation of important stress-related enzymes. Some nanomaterials can simulate the molecular patterns of specific pathogens without causing harm to the plant resulting in increased resistance to the disease. In the literature, chitosan, liposomes, and polysaccharide NPs have been reported to trigger plant immune responses that lead to the expression of defense-related genes and have been found to significantly reduce the severity of plant diseases caused by bacteria, fungi, and viruses [107, 108]. Plant viruses contribute significantly to agricultural losses, accounting for nearly 47% of crop damage. NMs show promise in combating several viruses like potato virus Y (PVY), cucumber mosaic virus (CMV), and bean yellow mosaic virus (BYMV). NPs serve as delivery vehicles for nucleic acids, sustaining plant immunity to develop resistance against viral infection via RNA interference (RNAi) [109]. Further, they act as standalone protectants or carriers for pesticides and RNA-interference compounds, while also exhibiting virucidal activity through mechanisms such as reactive oxygen species (ROS) production and interference with viral binding to manage plant diseases. Nanoparticles interfere with virus recognition by host cells through their interactions with virus surface proteins via glycoprotein receptors. For example, zinc oxide NPs (ZnONPs), iron oxide NPs (Fe3O4NPs), and Schiff-based nano-silver NPs have shown inhibitory effects against tobacco mosaic virus (TMV) infection [110]. Similarly, clay nanosheets loaded with plasmid DNA expressing artificial microRNAs (amiRNAs) have demonstrated efficacy against tomato yellow leaf curl virus [111]. Additionally, nanomaterials enhance the stability, translational efficiency, and cellular targeting of mRNA in plant genetic engineering, exemplified by BioClay-mediated protection against pepper mild mottle

Sr. no.	Nanomaterial	Pathogen	Activity	References
1	Ag NP	Xanthomonas campestris and X. axonopodis	Antimicrobial activity	[76]
2	Ag NPs	Bipolaris sorokiniana	Inhibited colony formation (in vitro)	[62]
3	Ag NP	Botrytis cinerea (grey mold)	Antifungal properties	[51]
4	Ag NPs	Alternaria alternata, Macrophomina phaseolina,	Strong antifungal activity	[77]
5	Ag NPs (solution)	Sphaerotheca fusca	Inhibited the fungus growth (in vitro and vivo)	[78]
6	Ag NPs	F. culmorum	Inhibited the fungus growth (in vitro)	[79]
7	Ag NPs	Xanthomonas oryzae pv. oryzae	Antibacterial activity	[80]
9	Ag NPs	Fusarium oxysporum	Fusarium wilt disease management	[81]
10	Ag NPs	Bean yellow mosaic virus	Mosaic disease	[82]
11	Ag NPs	Phytophthora arenaria	Crown and root rot disease inhibition	[83]
12	Ag NPs	Pyricularia grisea	Antifungal activity	[84]
14	Ag NPs	Rhizoctonia	Solani sheath blight	[85]
15	Ag NPs	Fusarium graminearum, F. sporotrichioides, F. avenaceum	Inhibited the development of fungus	[86]
18	Cu NPs	F. culmorum, F. oxysporum	Inhibited fungal growth	[87]
19	Cu NPs	Alternate, P. destructiva	Antifungal activity	[88]
20	Cu NPs	Penicillium digitatum	Green rot	[83]
21	CuO-NPs	Colletotrichum gloeoesporioides	Antifungal activity	[89]
22	ZnO NPs	Burkholderia glumae	Antibacterial activity	[90]
23	TiO ₂ NPs	Bipolaris sorghicola	Target leaf spot	[91]
24	TiO ₂ NPs	Dickeya dadantii	Antimicrobial agent	[92]
25	TiO ₂ NPs	Bipolaris sorokiniana	Spot blotch disease	[93]
26	SiO ₂ NPs	Aspergillus flavus	Reduced Ear rot disease	[91]
27	SiO ₂ NPs	Tobacco mosaic virus	Mosaic disease	[94]
28	Chitosan NPs	Xanthomonas, Erwinia strains	Inhibited the growth of both pathogens	[95]
29	Chitosan-magnesium NPs	A. oryzae and R. solani	Antimicrobial agent	[96]
30	CS NPs	Fusarium graminearum	Antifungal agent	[97]
32	Titanium dioxide (TiO ₂) NPs	wheat rust (Ustilago tritici)	Antifungal activity	[98]
33	Cu-chitosan	Leaf and stem rust of wheat	Inhibited to diseases	[99]
34	Chitosan	Alternaria leaf spot disease	Fungicidal properties	[44]
35	Chitosan	C. gelosporidies, G. fujikori, P. capsici, S. sclerotiorum, F. oxysporum	Antifungal activity	[95]
36	Chitosan NPs	Pyricularia oryzae	Antifungal activity	[100]
37	MgO NPs	Rhizoctonia solani; Acidovorax oryzae	Antimicrobial agent	[96]
38	MgO NPs	Xanthomonas oryzae oryzae	Antibacterial activities	[101]
39	Silicon dioxide NPs	Xanthomonas oryzae pv. oryzae (Xoo)	Antibacterial action	[102]
40	AI2O3 NPs	Fusarium root rot	Inhibited fungal growth	[103]
41	CWP-NPs	F. oxysporum f.sp. lycopersici	Antifungal activity	[104]
42	Se NPs	Phytophthora infestans	Antifungal activity, elicit resistance	[105]

Table 1 Application of nanomaterials for disease resistance in crops

virus (PMMoV) and CMV [112]. Furthermore, NMs target cell shapes, membrane integrity, essential biomolecules, enzymes, and pathogen-related proteins to exert inhibitory and anti-microbicidal effects. Scientists have postulated that NMs activate reactive oxygen species (ROS) and secondary signaling messengers that result in transcriptional regulation within plant secondary metabolism, but much work still needs to be done to clarify the mechanism [113]. Previously, scientists have used nanofibers, nano-capsules, and nanoparticles to successfully regulate gene expression. For example, the different changes in gene expression were studied by quantitative RT-PCR, and relative levels of expression of PR1, LoxA, Osm, and GluA were measured in roots and hypocotyls of plants at 12, 24, 72, and 120 h after treatment [114]. In the model plant *Arabidopsis thaliana*, ROS production condensed by Ag NPs lowered stress enzymes and induced autophagy. Multiple deformations on the spores of *A. brassicicola* were discovered using a scanning electron microscope [115]. Likewise, *Rhizoctonia solani*, *Macrophomina phaseolina*, and *Alternaria alternata* can all be inhibited by Cu-CS NPs [116].

PR proteins

Upon pathogen detection, plants rapidly mobilize diverse defense mechanisms to combat infection. Among these, the timely induction of pathogenesis-related (PR) proteins plays a pivotal role in establishing early resistance against invading microbial pathogens. This is initiated within the cell by the identification of pathogen effectors through plant resistance proteins, frequently the nucleotide binding site (NBS)-leucine rich repeat (LRR) proteins. Furthermore, to manage plant diseases, systemic acquired resistance (SAR) also uses a natural signaling pathway comprising SA, reactive oxygen species (ROS), and nitric oxide (NO). Salicylic acid (SA) contributes to SAR by activating genes involved in pathogenesis (PR) [117]. Similarly, resistance genes are expressed in plants after pathogen infection when nanoparticles (NMs) are applied. For example, silicon nanoparticles (SNPs) demonstrate efficacy in activating tomato plant defenses via systemic acquired resistance pathways, as evidenced by the upregulation of crucial pathogenesis-related and antioxidant genes upon application. Ultrastructural analysis reveals SNP distribution in plant tissues directly correlates with inhibition of in planta pathogens. SNPs mitigate reactive oxygen species, membrane damage, and pathogen growth, underscoring their potential as a sustainable bioprotectant to enhance crop resistance against infection through diverse complementary mechanisms [118]. Similarly, bacterial growth on Arabidopsis leaves was assessed to quantify local systemic resistance to a virulent strain of *P. syringae* under control conditions. SiO₂ NP and Si (OH)4 application revealed a mechanistic insight into the processes involved in the induced triggering of SAR [119].

Reactive oxygen species (ROS)

ROS, including hydrogen peroxide (H_2O_2) , superoxide (O_2^-) , and hydroxyl radicals $(OH \cdot)$, are highly reactive molecules generated as part of the plant's defense mechanisms when opposed by pathogenic microorganisms such as fungi, bacteria, and viruses. These ROS have the capability to harm cell membranes, proteins, and DNA, resulting in the pathogen's death [120]. The integration of nanomaterials in agriculture has led to a new era of innovative strategies to combat plant diseases and enhance crop productivity. In this context, the role of ROS in plant pathogen interactions has attracted substantial attention. Few nanomaterials can imitate the molecular patterns of pathogens and trigger the production of ROS in plants.

The production of ROS by these nanoparticles leads to an oxidative burst, which can cause damage to the pathogen's cell membrane and ultimately kill the pathogen [121]. Genetically, specific enzymes produce ROS under certain developmental or hormonal regulation to initiate or propagate signaling pathways [122]. For instance, the expression of defense-related genes, reactive oxygen species, and ATP-binding cassette (ABC) in wheat increased quickly after inoculation and pathogen attack. Similarly, ROS were activated during the application of NMs as the antimicrobial properties of graphene oxide nanoparticles were also observed, similar to pathogen membrane damage [123]. Conclusively, the production of ROS by nanomaterials can be an effective mechanism for fighting plant pathogens and can be an efficient way to reduce the use of chemical pesticides. However, the production of high levels of ROS can also be detrimental to the plant; therefore, it is important to find the optimal balance between activating the plant's defense mechanisms and not causing harm to the plant.

NMs for the induction of phytohormones and signaling molecules

Plants activate the immune system in response to pathogen attack, resulting in several physiological alterations in the plant body. The induction of immune responses is hypothesized to be regulated by phytohormones (jasmonic acid, methyl jasmonate, and salicylic acid) and signaling molecules (reactive oxygen species) [124]. Likewise, liposomes which are spherical formations consisting of a double layer of phospholipids have the capability to enclose and transport these molecules directly to plant cells. This action initiates the activation of genes associated with the plant defense system [68]. For instance, polymeric nanoparticles such as poly (lactic-co-glycolic acid) (PLGA) nanoparticles can be used to deliver phytohormones and signaling molecules to plants [125]. In addition, chitosan nanoparticles have been explored as a delivery vehicle for phytohormones and signaling molecules. These NPs can protect the delivered molecules from degradation and target them to specific cells in the plant leading to an enhanced defense mechanism as shown in Fig. 2.

Furthermore, the natural immune response is activated when pathogen-associated molecular patterns (PAMPs) are perceived by the host plants, which inhibits the development of plant infection. Plant hormones primarily control the events that affect how plants grow and develop. Abscisic acid (ABA), auxins, cytokinin (CK), gibberellins (GA), brassino steroids (BR), ethylene (ET), salicylic acid (SA), and peptide production have all been shown to vary in plants that are infected [127]. Similarly, the inhibition of the rice blast fungus *Pyricularia grisea*



Fig. 2 Nanomaterials activate defense responses in crops against pathogens. The diagram provides a clear illustration of the role of nanomaterials in activating immune responses and suppressing plant pathogens. One example of this is the use of SiO₂ NPs, as demonstrated in a recent study [126]. Nanomaterials work through various mechanisms in both plant and pathogen cells, including the activation of immune responses and the destruction of pathogen cells. One important mechanism through which nanomaterials activate immune responses in plants is through the induction of systemic acquired resistance (SAR). Elicitor application to plants can also activate signals to distant tissues, and salicylic acid (SA) is a plant hormone that plays a significant role in the initiation of SAR by activating pathogenesis-related (PR) genes. Nanomaterials can also be effective in destroying pathogen cells by regulating signaling pathways and inducing the production of salicylic acid and reactive oxygen species (ROS). This can help suppress the growth and spread of plant pathogens and prevent damage to crops

by chitosan nanoparticles is one example and some polymeric nanoparticles can also induce resistance against diverse plant pathogens. Changes in gene expression such as those for peroxidase, phenylalanine ammonia lyase, catalase, superoxide, and polyphenol oxidase were linked to greater resistance against pearl millet downy mildew following treatment of seeds with chitosan nanoparticles [128]. Copper and silicon nanomaterials can increase the amount of both enzymatic and non-enzymatic plant immune chemicals in tomato, leading to increased disease tolerance to Clavibacter michiganensis and ultimately increasing tomato crop performance [129]. Similarly, other research examined the antibacterial activity of magnesium oxide nanoparticles (MgO NPs) and its implications on disease resistance in tomato plants against Ralstonia solanacearum. After treatment with MgO NPs, tomato plant roots and hypocotyls showed increased levels of salicylic acid-inducible PR1, jasmonate LoxA, and systemic resistance-related GluA [114].

Defense-related enzymes

Molecular biologists stated that proteins perform all the tasks necessary for crop development, maturity, and immunity. Specific enzymes and peptides have antimicrobial properties. The researchers investigated defense mechanisms at the biochemical, cellular, and transcriptomic levels. Transcriptional analysis found mycogenic SeNPs upregulated important genes related to phenylalanine lyase, lipoxygenase, β -1,3-glucanase and superoxide dismutase, correlated with increased enzymatic activities important for biochemical defenses. Treated plants also accumulated significantly higher levels of important cellular defense molecules like callose, lignin and hydrogen peroxide compared to controls. These findings provided mechanistic insights showing mycogenic SeNPs activate

robust biochemical and molecular defenses in tomatoes to combat late blight infection. Enzymatic and nonenzymatic anti-oxidative defense mechanisms are both activated in plants and combine to eliminate free radicals [130]. For instance, research has shown that silver NPs can induce the production of enzymes such as peroxidases and catalases in plants which can help to produce resistance against pathogens by breaking down harmful molecules and reducing oxidative stress. Similarly, NMs enhanced plant growth and boost the levels of self-protective enzymes including super oxide dismutase, CAT (catalase), and phenylalanine ammonia lyase (PAL), respectively [131]. The application of chitosan-based nanomaterials to maize improves resistance against plant pathogens by modifying ROS-scavenging enzymes such as CAT, peroxidase POD, and SOD [132]. Phenylalanine ammonia lyase is an important enzyme that produces antimicrobial molecules (like phytoalexins and pathogenrelated proteins) and facilitates colonization around the infection point in plants [133]. The levels of lignin, callose, and hydrogen peroxide that act as the cells defense in the primed plants recorded a significant increase above the control plants. Similarly, Se NPs can be employed as a nano-bio stimulant antifungal to tomato plants by triggering immune responses against tomato late blight [105]. It has been reported by [134] that tomato-treated plants using chitosan nanoparticles increased the expression of SOD and CAT and protected the plants against bacterial wilt disease. Similarly, Sathiyabama and Indhumathi [135] conducted the latest study to determine how chitosan thiamine nanoparticles (TC NPs) affected the activation of innate immunity in chickpeas against stress brought on by the wilt pathogen Fusarium oxysporum f. sp. ciceri (FOC) under greenhouse conditions. In plants treated with TC NPs, there was more than 90% wilt resistance. In TC NPs-treated plants, histochemical staining revealed significant lignin development in the vascular bundles of chickpea stem cells. Several nanomaterials that induce immunity against plant pathogens are given in Table 2.

Nanomaterials as diagnostic tools for crop plant diseases

Identification of plant pathogens and disease incidence is important for managing plant disease and infestation. It allows for the observation and management of disease infections at different phases of the disease development cycle in open field and greenhouse studies. Plant pathogens have been widely detected using a variety of approaches including biomarkers, immunoassays, serological tests, and DNA-based techniques [145]. Hence, remote sensing tools are valuable for disease detection in plants. The core concept of remote sensing involves the use of non-contact, regularly monitoring instruments such as infrared red, chlorophyll fluorescence detection, and 3D scanning to collect information regarding activities occurring in both natural and human-engineered environments. For example, in a laboratory setting, Bawden noticed in 1933 the dramatic differences seen between necrotic leaf patches created by potato and tobacco viruses in an infrared image. The necrotic cells in potatoes contained chemical breakdown products, whereas the necrotic cells in tobacco were empty and differed in color from healthy leaf cells. These discoveries laid the foundation for the use of various spectral bands to identify variations in plant health [146]. Nanomaterials such as nanoparticles and nanocomposites have potential applications as diagnostic tools for plant disease

Tab	e 2	App	lication	of r	nanoma	iterials	that	induce	e resistan	ce to	crop p	plant p	bathogens
-----	-----	-----	----------	------	--------	----------	------	--------	------------	-------	--------	---------	-----------

Sr. no.	Nanomaterial	Pathogen	Activity	References
1	Chitosan oligosaccharide nanoparticles (Cos-La)	Magnaporthe grisea	Defense response against rice blast	[136]
2	Fe ₃ O ₄ NPs	TMV	Resistance against TMV through upregulation of SA genes	[110]
3	ZnO NPs	TMV	Deactivation of TMV particles and regulation of plant immunity	[137]
4	NiO NPs	Cucumber mosaic virus CMV	Reduced virus titer	[138]
5	Ag NPs	PVY, ToMV	Induce SAR, suppress their infection on tomato plants	[139]
6	Ag NPs	Cucumis sativus (crop)	Activated antioxidant defense systems	[140]
7	CuO NPs	Lens culinaris (Crop)	Defense enzyme activator	[141]
8	ZnO NPs	Tomato mosaic virus (ToMV)	Induce defense system activity	[142]
9	Elemental Sulfur NPs	Fusarium oxysporum f. sp. lycopersici	Salicylic acid-dependent systemic acquired resistance	[118]
10	Carbon-based NPs	Fusarium wilt in tomato	Antioxidant defense system	[143]
11	CuFe NPs	Meloidogyne incognita and Meloidogyne	Nematicide activity	[144]

identification. For instance, gold nanoparticles have also been explored as a potential tool for detecting specific plant pathogens [147]. Gold nanoparticles have also been employed for the detection of Begomovirus in chili and tomato plants, with the capability to detect 500 ag/µL of begomoviral DNA [148]. Furthermore, researchers have used this approach to detect specific plant pathogens such as *Phytophthora infestans*, a fungal pathogen that causes potato and tomato late blight, and Xanthomonas campestris pv. vesicatoria, a bacterium that causes bacterial spot disease in pepper and tomato. Moreover, nanobiosensors are very helpful such as molecular assays and smartphone apps for plant pathogen detection and disease monitoring in comparison with traditional methods [149]. Similarly, remote sensing techniques are used because plant infections and pest attacks normally alter how light interacts with leaves and branches. The use of nanosensors in plant quarantine and seed certification may prove to be an efficient and precise method for the identification of pathogen infections in plants. Therefore, the high sensitivity and specificity of NM-based biosensors, which are essential for the early diagnosis of plant pathogens, are just a few of their many advantages over conventional biosensors [150]. However, nano-biosensors are designed to interact with biological entities such as proteins, nucleic acids, cells, or even whole organisms enabling the detection of various biochemical processes, biomarkers, or pathogens [151]. These sensors revolutionize agriculture by swiftly detecting plant pathogens through nanomaterials and bioreceptors, enhancing accuracy. Nano-biosensors exhibit remarkable sensitivity detecting even trace amounts of pathogens, thereby aiding in early disease diagnosis. They are more useful for smart agriculture because of their low detection limit and high sensitivity [152]. The agricultural land could be monitored in real time using a nano-biosensor with a Global Positioning System (GPS). This technique allows for the early detection of the pathogen and information on crop growth. For example, a study reported that surface plasmon resonance can be used by nano-gold-based immunosensors to identify the pathogen Tilletia indica that causes Karnal bunt in wheat [153]. Similarly, gold NPs are used in nano-biosensors because of their transiting nature between optical and electrochemical methods for pathogen identification [154]. Furthermore, a metalloporphyrin-based e-nose offers a novel approach for accurately identifying and tracking Fusarium-infected wheat grains. Metalloporphyrins, as complex molecules with metal ions exhibit specific interactions with volatile organic compounds (VOCs) released by plants during stressful conditions including those generated as a response to pathogenic infections [154]. However, in plant tissue culture different NPs including ZnO NPs, ${\rm TiO}_2$ NPs, and Ag NPs, are primarily utilized to limit microbial activity.

Electronic nose (e-nose) system

The relationship between electronic noses and nanomaterials lies in the development and improvement of sensors used in electronic nose systems. Nanomaterials can be employed in various ways to enhance the performance of e-nose sensors. A sensor-based intelligent device called an "e-nose system" is created to recognize and classify complex scents using various non-selective sensors. The electronic nose (e-nose) system is inspired by the human olfactory system, which has diverse applications spanning food quality assessment, environmental monitoring, and safety enhancement. In the context of plant pathology, e-noses offer a rapid and non-destructive approach for the early detection of plant pathogens. By using an array of gas sensors, these systems mimic human olfaction and detect volatile organic compounds (VOCs) emitted by plants in response to pathogen infections. This technology has the potential to revolutionize plant disease management by enabling proactive measures against the spread of diseases and crop losses [155]. For example, an e-nose system was developed to detect and differentiate between fungal pathogens affecting wheat crops based on the distinct VOC profiles emitted by each pathogen. The results demonstrated the system's ability to accurately classify infected wheat samples and identify the specific pathogens responsible for the infections. Another study focused on the use of an e-nose for the early detection of bacterial canker disease in tomato plants. By analyzing the VOCs released by infected plants, the e-nose system successfully discriminated between healthy and infected tomato samples, demonstrating its potential as an efficient diagnostic tool [156]. Similarly, E-noses are a reliable method for identifying fungal infections on rice grains before visible symptoms appear. Jiarpinijnun et al. [157] used to detect Aspergillus fungus presence on Jasmine brown rice grains. Furthermore, it has shown promise in real-world agricultural settings. An e-nose system was deployed to monitor and identify the presence of the fungus Botrytis cinerea in grapevine crops [158]. The system's accuracy in detecting the pathogen was demonstrated through its ability to detect infected plants even before visible symptoms were apparent, facilitating timely interventions to control the disease and minimize crop losses. Through its ability to accurately detect VOCs emitted by infected plants, e-nose offers a non-invasive, early detection approach that can aid in reducing the spread of plant pathogens and enhancing agricultural productivity.

Moreover, the use of specifically designed nano-biosensors can be developed to contribute to sustainable agriculture through the reduction of plant pathogens. A biosensor is designed to detect pathogens and signals when control measures are required. However, it does not possess the capability to directly control or mitigate the pathogens themselves. In addition, hyperspectral sensors streamline daily tasks by enabling them to be performed on smartphones, enhancing efficiency. Unlike conventional methods that require extensive field monitoring, farmers can now assess crops more efficiently using hyperspectral sensors. Farmers can effectively collect information such as the spectral signature of crops and interact with agricultural organizations to run their land from their homes [159]. A smartphone application called Dr. Lada was created in Malaysia by scientists at the University Kebangsaan Malaysia [160]. This program was employed to identify pathogens and pests in pepper. By enabling early detection of plant pathogens, these sensors can help prevent the spread of disease and reduce the economic losses associated with crop damage. Furthermore, pre-symptomatic and disease-specific identification and the impact on the environment remain significant issues in the electronic monitoring of plant pathology. By responding to questions, users could determine whether a pest or disease infection was present which reduced the need for farmers to rely on agricultural officers and allowed them to identify diseases on their own. Furthermore, it should be emphasized that managing farmers' demands must be the primary objective of electronic plant pathology. However, more research is needed to fully understand the potential of nanomaterials as diagnostic tools for plant disease identification and to ensure their safe and effective use in agriculture (Fig. 3).

Direct applications of nanomaterials against crop pathogens

The accurate method of direct application of nanomaterials against pathogens remains unclear. Usually, leaves, roots, and other vegetative portions of different plants absorb nanomaterials. NMs enter the plant via natural opening sites such as stomata, hydathodes, stigmas, and wounds [164]. Many assumptions on how nanomaterials work is consistent with their distinctive physicochemical characteristics, such as size, surface-to-volume ratio, and shape. NMs of smaller sizes have a greater surface area per unit volume which increases the possibility that they meet bacteria, viruses, and fungi which can cause cell death and damage. The exact mechanism of action of nanomaterials against plant pathogens can vary depending on the specific nanomaterial, pathogen type, and environmental conditions. However, many nanomaterials such as silver, zinc oxide, and titanium dioxide NPs can generate reactive oxygen species (ROS) when exposed to light or other forms of energy. These ROS can damage the cell membrane and DNA of pathogens leading to their death [165]. Similarly, chitosan nanoparticles can inhibit the activity of enzymes essential for the survival of pathogens. Nanoparticles can interfere with the metabolic pathways of pathogens, which makes it difficult for the pathogens to survive. Several studies have been reported and the in vitro antiviral activity of SiO₂ NPs and ZnO NPs against TMV was revealed in an investigation. It has been hypothesized that NPs could directly interact with viral capsid proteins leading to structural distortion that causes TMV aggregation and rapid viral particle inactivation [110]. In the case of viruses, nanoparticles (NPs) exhibit direct antiviral effects by interfering with viral genome replication and protein biosynthesis, particularly viral coat protein. They impede viral genome packaging and capsid protein degradation crucial for virus particle assembly. Additionally, NPs may disrupt plant cell electron transport systems, enhance cellular barriers, block viral entry, and inhibit viral DNA replication [109]. Moreover, little is known about the temporal changes in infection and efficient treatment plans. Each of these elements must be understood at a systematic and molecular level to fully investigate and recognize the potential of NM management techniques. It is also important to appropriately evaluate NPs including their placement, dosage, and timing to effectively counter pathogens. The use of nanoparticles in plant pathology is still under research and not widely used in commercial agriculture.

Biosafety perspective

The use of nanomaterials (NMs) against plant pathogens raises several concerns from a biosafety perspective. To fill the information gap about the hazardous consequences of NMs when they enter a different plant species, a deeper understanding of the biosafety issues is mandatory. For instance, particle size significantly impacts their physical and chemical characteristics as smaller nanoparticles (<10 nm), higher concentrations, and specific shapes induce greater phytotoxic effects, contrasting with larger, lower-concentration, or more spherical nanoparticles [166, 167]. Variability in plant species sensitivity to nanoparticle exposure is observed; titanium dioxide NPs triggered higher reactive oxygen species production and DNA damage in wheat roots compared to soybean [168]. Conversely, zinc oxide NPs inhibited rice shoot and root growth more profoundly than cucumber, with cucumber displaying enhanced antioxidant activity against NPinduced oxidative stress [169]. However, the main use of nanomaterials is to reduce the need for agrochemicals fertilizers and fungicides while increasing production through effective management of plant pathogens and pests, though frequent use of fungicides can cause



Fig. 3 Nanomaterials as diagnostic tools. This illustration describes the process of plant pathogen identification using electronic nano-biosensors and nanomaterial-based sensors. In the first stage, sensing components are used to detect pathogenic molecules in plant tissues. These sensing components are typically electronic devices that are functionalized with biological recognition elements, such as plantibodies^{*}, that can specifically bind to pathogenic molecules. When the pathogenic molecule binds to the recognition element, it triggers a signal that can be detected by the electronic device. In the second stage, the scanning elements typically consist of nanomaterial-based scanning elements, such as carbon nanotubes or gold nanoparticles, which are functionalized with biological recognition elements. When pathogenic cells or spores meet the nanomaterials, they bind to the recognition element on the nanomaterials. This binding between the pathogenic cells and the nanomaterials triggers a change in the nanomaterials themselves. These changes can be their chemical properties, such as color or conductivity. Here is a highly sensitive device designed (nano-sensor) to detect even the smallest changes at the nanoscale. In the final stage, data analysis is used to interpret the signals generated by the sensing and scanning elements. This involves the use of algorithms and machine learning techniques to analyze the data and identify the specific pathogens present. (* The concept of plant-produced antibodies, commonly known as "plantibodies", was first demonstrated by Hiatt and Duering in 1990 [161]. This term is used to describe antibodies produced and expressed in plants [162]. It can be used for various purposes, including pathogen detection, and immunizing the plant against pathogen infection [163])

environmental contamination and pathogen resistance. Further, alterations in microbial diversity and activity can disrupt nutrient cycling, soil fertility, and plant-microbe interactions, compromising crop health. Soil microbes, essential for nutrient cycling and disease suppression, may be adversely affected by nanomaterial exposure, leaving crops more vulnerable to pathogens [18, 19]. Also, some engineered nanomaterials, like silver and zinc oxide NPs exhibit antimicrobial properties, causes microbial imbalances which can lead to increased susceptibility to diseases in crops by affecting plant growth-promoting rhizobacteria (PGPR) and mycorrhizal fungi [170, 171]. The coexistence of natural and engineered nanomaterials in soil poses contamination risks, with highly produced ENMs like TiO_2 , ZnO potentially surpassing toxic thresholds [172]. Consequently, nanomaterial presence alters the plant rhizosphere, affecting microbial communities, enzyme activity and plant health. Studies indicate toxicity in rice plants [14, 173] and microbial communities exposed to nanomaterials, underscoring the need for cautious agricultural nanomaterial use. Therefore, greensynthesized NMs are needed to produce eco-friendly alternatives, such as chitosan, a natural polymer that has been shown to be an effective substitute for defending hosts from fungal infections [174]. For instance, it has been recommended that chitosan may increase the

expression of 1,3-glucanase and chitinase genes in addition to the activity of defense-related enzymes in tomato and Arabidopsis [175]. Furthermore, nano-based agricultural products play a crucial role in managing plant pathogens as discussed in Table 1 and 2. Their appropriate handling, long-term storage capability, ease of transport, non-toxic nature, and high effectiveness position them as an ideal choice for farmers compared to traditional chemicals [176]. Nano-based chemicals are rapidly gaining global interest and seeing significant growth in commercialization. Similarly, commercial nano-formulations are increasingly employed in agriculture. Nano-Ag Answers® (Urth Agriculture) utilizes silver nanoparticles as a potent biocide against predatory pests. Nano-GroTM (agro nanotechnology corporation) incorporates nanomaterials to stimulate plant immune system pathways and boost defense mechanisms. ZinkicideTM, employing zinc nanoparticles, demonstrates effectiveness in managing citrus scab, canker, and melanoses [91]. In addition to these developments, various regulatory laws and policies have been established in the field of nanotechnology. A noteworthy achievement in 1998 was the national science and technology council's initiative which resulted in the creation of the national nanotechnology initiative (NNI). The primary goal of NNI was to conduct research and development to address and build awareness about nanobased products within the community [91]. However, a scientific advisory panel under the federal insecticide, fungicide, and rodenticide act in the United States has raised concerns about the environmental risks posed by nano-silver oxides in pesticides. The panel suggests that there is inadequate potential to mitigate these effects. In Europe, comprehensive regulatory frameworks both vertical and horizontal have been established to screen and manage the risks associated with nanoparticles (NPs) to both the public and the ecosystem. Moreover, plan protection products (PPPs) are primarily regulated by rule (EC: 1107/2009), necessitating previous agreement before being introduced to the commercial market [177]. The authorization process involves two steps, with the European Food Safety Authority (EFSA) analyzing the operating parts used in PPPs, and European member states screening and approving the product nationwide. However, regulatory frameworks and policies in this regard are yet to be developed [178]. Additionally, comprehensive analytic tools perform an important role in the evaluation of regulatory laws for risk assessment. To manage pesticide resistance, there is a need for the rotation of pesticide groups to prevent the emergence of new strains. Also, a diverse array of nanopesticides is anticipated to be commercially available in the coming years. However, the development of nanoproducts faces challenges due to a limited understanding of their performance in field trials,

high production costs, significant volume demands, and concerns regarding regulatory and public perceptions [179]. However, it is still necessary to solve their drawbacks in terms of price, preparation, particle dispersion, and ingredient delivery.

Nanomaterial challenges against crop plant pathogens

The risk assessment of nanomaterials in crop health is pivotal for ensuring safe agricultural practices. Currently, there are no standardized methods or regulations for assessing the toxicity and safety of nanomaterial-based agrochemicals. For instance, nanoparticle-induced phytotoxicity was evaluated using seeds of various crops, including Allium cepa, Zea mays, Cucumis sativus, and Lycopersicum esculentum [180]. Furthermore, regulatory frameworks must be established to oversee nanomaterial usage, ensuring adherence to safety protocols [181]. In vitro studies underscore the cytotoxic effects on seeds and seedlings such as mitotic index changes, chromosomal aberrations, and DNA damage, highlighting the imperative for comprehensive environmental risk assessments in crops [182]. It is evident from the literature that nanomaterials could be used for years for plant disease monitoring and resistance enhancement in crop plants to ensure food security. However, there are always objections regarding the adoption of new technology; hence, further in-depth research on their implications is urgently needed. Few hazardous effects of NMs may vary depending on the bulk substance, particle size, and dose employed to create them. Similarly, several studies found that exposure to single-walled carbon nanotubes, ZnO NPs, Ag NPs, and Fe nanomaterials led to reduced seed germination and downregulated gene expression in wheat, maize, barley, ryegrass, and soybeans. Likewise, increased silver ion concentrations can damage DNA and limit its ability to replicate. This results in the inactivation of ribosomal subunit proteins as well as other cellular proteins and enzymes that are required for ATP generation. Ag NPs have been proven in previous studies to have powerful antifungal effects on fungi by destroying membrane integrity [183]. Correspondingly, the use of NMs to develop plant disease resistance is a difficult task. Plant growth and development may be influenced by NMs, although their absorption, transport mobilization and targets in plant tissues are still poorly understood. At some doses, NPs may cause toxicities by altering the physiological and morpho-anatomical genetic components of plants [184]. The vacuole, apoplast, phloem tissues, and xylem of plant roots and shoots are among the sites where nanoparticles can be absorbed, dispersed, and accumulated. The use of nanomaterials (NMs) in crop plant pathogens is still an emerging field, and there

are several limitations to their use. NMs can have potential toxicity to plants, and their long-term effects on the environment are not yet fully understood. However, the use of green synthesis techniques for nanoparticle production could be a cost-effective and ecologically favorable alternative.

Concluding remarks and future perspectives

This article pioneers the exploration of this exciting frontier, envisioning a future where nanomaterials exhibit diverse biocidal activities, including fungicidal, bactericidal, and virucidal properties, among others. This has the potential to revolutionize disease resistance for crop pathogens, both in vivo and in vitro, indicating a new era of crop protection. Nanomaterials have the potential to revolutionize agriculture and lead to breakthroughs in plant disease management. They can stimulate plant immunity and inhibit the growth of pathogens by producing antimicrobial compounds and secondary metabolites. However, the use of nanotechnology in food and agriculture is still in its promising stage, and certain faults and risks exist such as phytotoxic behavior which needs to be thoroughly understood and determined at different plant growth stages. Despite these challenges, the use of nanomaterials in plant disease management can provide several advantages over traditional methods and can be tailored to target specific pathogens. Nanomaterials' potential applications throughout the food chain from farming to packaging are getting attention due to their unique properties. These properties provide opportunities to enhance food quality, safety, and sustainability. In addition, crop diseases threaten plant growth by disrupting biochemical and molecular processes, yet NMs offer significant potential in enhancing plant performance and resistance to pathogens. NMs improve membrane stability, nutrient uptake, and protect photosynthetic apparatus from pathogen damage. They also enhance the accumulation of stress-protective compounds and upregulate stress-responsive genes, boosting plant defense mechanisms. Recent research extensively explores NMs' role in inducing tolerance to crop pathogens. However, investigations into the influence of NMs on proteomics and genetic factors remain limited at molecular level, highlighting the need for further investigation in future studies to better understand these aspects. Further research and regulation will be necessary to ensure the safe and effective use of nanomaterials in agriculture. In terms of future policies, nanotechnology shows promise in addressing various challenges related to plant characteristics, productivity, and resistance to pathogens. However, concerns regarding their long-term ecological impacts, including bioaccumulation in food chains and toxicity to environmental organisms, necessitate thorough environmental risk assessments. Integrating green synthesis methods for nanoparticles can mitigate their toxicity. Biogenically synthesized nanoparticles exhibit enhanced bioactivity against pathogenic microbes, surpassing their chemically synthesized counterparts [185]. Additionally, diverse nanomaterials, such as nano clays and nanotubes, offer unique properties like enhanced sensitivity and rapid response times, with polymer-based nanocarriers such as silica and chitosan serving as protective reservoirs for encapsulating pesticides [186]. In addition, nano-sensors and nano-devices are emerging as innovative tools for real-time pathogen monitoring in plants with potential applications spanning pre- and post-infection detection under both laboratory and field conditions. This surge in interest agrees with the rapid growth of the NMs industry, fueled by large-scale production and heightened demand for NMs-derived products like nanoscale carriers and biosensors. However, unlocking the full potential of nano-agrochemicals requires a balanced approach of responsible exploitation, stringent regulatory frameworks, and continuous monitoring.

Though, it is important to thoroughly examine how nanomaterials interact with crops and assess their ecological and toxicological impacts before considering their implementation, including the assessment of gene expression patterns. Combining nanomaterials with other disease management strategies can also lead to synergistic effects. Despite existing applications, the vast potential of NMs in plant protection continues to lie dormant. Emerging research on their antimicrobial capabilities highlights their immense promise for revolutionizing crop health through novel diagnostic and sustainable management solutions.

Abbreviations

ZnO NPsZinc oxide nanoparticlesCuO NPsCopper oxide nanoparticlesZnO NPsZinc oxide nanoparticlesTiO2 NPsTitanium dioxide nanoparticlesSiO2 NPsSilicon dioxide nanoparticlesCS NPsChitosan nanoparticlesS NPsSilicon nanoparticlesCuCh NPsCu-chitosan nanoparticlesMgO NPsMagnesium oxide nanoparticlesAl2O3 NPsAluminum oxide nanoparticlesCWP NPs (chitosan)Cell wall polymer-based nanoparticlesSe NPsMycogenic selenium nanoparticles
CuO NPsCopper oxide nanoparticlesZnO NPsZinc oxide nanoparticlesTiO2 NPsTitanium dioxide nanoparticlesSiO2 NPsSilicon dioxide nanoparticlesCS NPsChitosan nanoparticlesS NPsSilicon nanoparticlesCuCh NPsCu-chitosan nanoparticlesMgO NPsMagnesium oxide nanoparticlesAl2O3 NPsAluminum oxide nanoparticlesCWP NPs (chitosan)Cell wall polymer-based nanoparticlesSe NPsMycogenic selenium nanoparticles
ZnO NPs Zinc oxide nanoparticles TiO2 NPs Titanium dioxide nanoparticles SiO2 NPs Silicon dioxide nanoparticles CS NPs Chitosan nanoparticles S NPs Silicon nanoparticles CuCh NPs Cu-chitosan nanoparticles MgO NPs Magnesium oxide nanoparticles Al ₂ O ₃ NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
TiO2 NPs Titanium dioxide nanoparticles SiO2 NPs Silicon dioxide nanoparticles CS NPs Chitosan nanoparticles S NPs Silicon nanoparticles CuCh NPs Cu-chitosan nanoparticle MgO NPs Magnesium oxide nanoparticles Al ₂ O ₃ NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
SiO2 NPs Silicon dioxide nanoparticles CS NPs Chitosan nanoparticles S NPs Silicon nanoparticles CuCh NPs Cu-chitosan nanoparticle MgO NPs Magnesium oxide nanoparticles Al2 ₀ 3 NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
CS NPsChitosan nanoparticlesS NPsSilicon nanoparticlesCuCh NPsCu-chitosan nanoparticleMgO NPsMagnesium oxide nanoparticlesAl ₂ O ₃ NPsAluminum oxide nanoparticlesCWP NPs (chitosan)Cell wall polymer-based nanoparticlesSe NPsMycogenic selenium nanoparticles
S NPs Silicon nanoparticles CuCh NPs Cu-chitosan nanoparticle MgO NPs Magnesium oxide nanoparticles Al ₂ O ₃ NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
CuCh NPs Cu-chitosan nanoparticle MgO NPs Magnesium oxide nanoparticles Al ₂ O ₃ NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
MgO NPs Magnesium oxide nanoparticles Al ₂ O ₃ NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
Al ₂ O ₃ NPs Aluminum oxide nanoparticles CWP NPs (chitosan) Cell wall polymer-based nanoparticles Se NPs Mycogenic selenium nanoparticles
CWP NPs (chitosan)Cell wall polymer-based nanoparticlesSe NPsMycogenic selenium nanoparticles
Se NPs Mycogenic selenium nanoparticles
Fe ₃ O ₄ NPs Iron oxide nanoparticles
NiO NPs Nickel oxide nanoparticles
CeO ₂ NPs Cerium oxide nanoparticles
CuFe NPs Copper/iron nanoparticles
TC NPs Chitosan thiamine nanoparticles
ROS Reactive oxygen species
PR Pathogen related proteins
SAR Systematic acquired resistance
TMV Tobacco mosaic virus
CMV Cucumber mosaic virus

TSWV	Tomato spotted wilt virus
PVY	Potato virus Y
ToMV	Tomato mosaic virus

Author contributions

M.J. wrote the manuscript and worked on figures. M.A.A. and A.Z. reviewed the manuscript. F.A. and G.M.D. contributed to literature search. S.H. and M.I. carried out reference searching. S.M.: data curation and investigation. L.G. funded, supervised, edited, and approved the final manuscript.

Funding

Xinjiang Major Science and Technology Projects (Research, development, and demonstration of key technologies for the green control of major pests on special and superiority crops in Xinjiang, 2023A02009).

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹State Key Laboratory for Biology of Plant Disease and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China.²Institute of Plant Protection, Xinjiang Academy of Agricultural Sciences / Key Laboratory of IntegratedPest Management on Crop in Northwestern Oasis, Ministry of P.R.China, Xinjiang Urumqi 830091, China. ³Department of Plant Pathology, University of Agriculture, Faisalabad 38000, Pakistan. ⁴Department of Microbiology, Government College University, Faisalabad 38000, Pakistan. ⁵Department of Biotechnology, Chonnam National University Yeosu, Chonnam 59626, Republic of Korea. ⁶Key Laboratory of Agro-Products and Safety Control in Storage and Transport Process, Institute of Food Science and Technology, Chinese Academy of Agriculture Sciences Beijing, Beijing, China. ⁷State Key Laboratory of Rice Biology and Ministry of Agriculture Key Laboratory of Molecular Biology of Crop Pathogens and Insects and Key Laboratory of Biology of Crop Pathogens and Insects of Zhejiang Province, Institute of Biotechnology, Zhejiang University, Hangzhou 310058, China.

Received: 8 March 2024 Accepted: 2 May 2024 Published online: 23 May 2024

References

- 1. Omran BA, Baek KH. Control of phytopathogens using sustainable biogenic nanomaterials: recent perspectives, ecological safety, and challenging gaps. J Clean Prod. 2022;372: 133729.
- Minoli S, Jägermeyr J, Asseng S, Urfels A, Müller C. Global crop yields can be lifted by timely adaptation of growing periods to climate change. Nat Commun. 2022;13:7079. https://doi.org/10.1038/ s41467-022-34411-5.
- Cucak M, Sparks A, de Moral RA, Kildea S, Lambkin K, Fealy R. Evaluation of the 'Irish Rules': the potato late blight forecasting model and its operational use in the Republic of Ireland. Agronomy. 2019;9:515.
- Farber C, Mahnke M, Sanchez L, Kurouski D. Advanced spectroscopic techniques for plant disease diagnostics. A review. TrAC Trends Anal Chem. 2019;118:43–9.
- Gust AA, Pruitt R, Nürnberger T. Sensing danger: key to activating plant immunity. Trends Plant Sci. 2017;22:779–91.
- 6. Lucas JA. Advances in plant disease and pest management. J Agric Sci. 2011;149:91–114.

- Hassanisaadi M, Bonjar AHS, Rahdar A, Varma RS, Ajalli N, Pandey S. Eco-friendly biosynthesis of silver nanoparticles using *Aloysia citrodora* leaf extract and evaluations of their bioactivities. Mater Today Commun. 2022;33: 104183.
- Girado JP, Wu H, Newkirk GM, Kruss S. Nanobiotechnology approaches for engineering smart plant sensors. Nat Nanotechnol. 2019;14:541–53.
- Bombo AB, Pereira AES, Lusa MG, de Medeiros Oliveira E, de Oliveira JL, Campos EVR, de Jesus MB, Oliveira HC, Fraceto LF, Mayer JLS. A mechanistic view of interactions of a nanoherbicide with target organism. J Agric Food Chem. 2019;67:4453–62.
- Agrawal S, Kumar V, Kumar S, Shahi SK. Plant development and crop protection using phytonanotechnology: a new window for sustainable agriculture. Chemosphere. 2022;299: 134465.
- Okey-Onyesolu CF, Hassanisaadi M, Bilal M, Barani M, Rahdar A, Iqbal J, Kyzas GZ. Nanomaterials as nanofertilizers and nanopesticides: an overview. ChemistrySelect. 2021;6(33):8645–63.
- Rajput VD, Singh A, Minkina T, Rawat S, Mandzhieva S, Sushkova S, Shuvaeva V, Nazarenko O, Rajput P, Verma KK. Nano-enabled products: challenges and opportunities for sustainable agriculture. Plants. 2021;10:2727.
- Safdar M, Kim W, Park S, Gwon Y, Kim Y-O, Kim J. Engineering plants with carbon nanotubes: a sustainable agriculture approach. J Nanobiotechnology. 2022;20:1–30.
- Hao Y, Yuan W, Ma C, White JC, Zhang Z, Adeel M, Zhou T, Rui Y, Xing B. Engineered nanomaterials suppress turnip mosaic virus infection in tobacco (*Nicotiana benthamiana*). Environ Sci Nano. 2018;5:1685–93.
- Dziergowska K, Michalak I. The role of nanoparticles in sustainable agriculture. In smart agrochemical for sustainable agriculture. Elsevier; 2022. p. 225–78.
- Farooq T, Adeel M, He Z, Umar M, Shakoor N, da Silva W, Elmer W, White JC, Rui Y. Nanotechnology and plant viruses: an emerging disease management approach for resistant pathogens. ACS Nano. 2021;15:6030–7.
- Rasheed A, Li H, Tahir MM, Mahmood A, Nawaz M, Shah AN, et al. The role of nanoparticles in plant biochemical, physiological, and molecular responses under drought stress: a review. Front Plant Sci. 2022;13: 976179.
- Zhang W, Jia X, Chen S, Wang J, Ji R, Zhao L. Response of soil microbial communities to engineered nanomaterials in presence of maize (*Zea* mays L.) plants. Environ Pollut. 2020;267:115608.
- Zhang Y, Liu N, Wang W, Sun J, Zhu L. Photosynthesis and related metabolic mechanism of promoted rice (*Oryza sativa* L.) growth by TiO2 nanoparticles. Front Environ Sci Engin. 2020;14:1–12. https://doi.org/10. 1007/s11783-020-1282-5.
- Mohammadi H, Esmailpour M, Gheranpaye A. Effects of TiO2 nanoparticles and water-deficit stress on morpho-physiological characteristics of dragonhead (*Dracocephalum moldavica* L.) plants. Acta Agricul Slo. 2016;107:385–96. https://doi.org/10.14720/aas.2016.107.2.11.
- Venkatachalam P, Priyanka N, Manikandan K, Ganeshbabu I, Indiraarulselvi P, Geetha N, et al. Enhanced plant growth promoting role of phycomolecules coated zinc oxide nanoparticles with p supplementation in cotton (*Gossypium hirsutum* L). Plant Physiol Biochem. 2017;110:118– 27. https://doi.org/10.1016/j.plaphy.2016.09.004.
- Farias IAP, Santos dos CCL, Sampaio FC. Antimicrobial activity of cerium oxide nanoparticles on opportunistic microorganisms: a systematic review. Biomed Res Int. 2018. https://doi.org/10.1155/2018/1923606.
- Yadav A, Yadav K, Ahmad R, Abd-Elsalam KA. Emerging frontiers in nanotechnology for precision agriculture: advancements. Hurdles Prospects Agrochem. 2023;2:220–56.
- Jha A, Pathania D, Sonu, Damathia B, Raizada P, Rustagi S, Singh P, Rani GM, Chaudhary V. Panorama of biogenic nano-fertilizers: a road to sustainable agriculture. Environ Res. 2023;235:116456. https://doi.org/ 10.1016/j.envres.2023.116456.
- Alghuthaymi MA, Rajkuberan C, Rajiv P, Kalia A, Bhardwaj K, Bhardwaj P, Abd-Elsalam KA, Valis M, Kuca K. Nanohybrid antifungals for control of plant diseases: current status and future perspectives. J Fungi. 2021;7:48. https://doi.org/10.3390/jof7010048.
- Hassanisaadi M, Chaichi M, Mirzaei S, Heydari M. Myconanoparticles: synthesis and probable role in plant pathogen management. In: Biotic stress management of crop plants using nanomaterials. CRC Press; 2023. p. 125–72.

- Baig N, Kammakakam I, Falath W. Nanomaterials: a review of synthesis methods, properties, recent progress, and challenges. Mater Adv. 2021;2:1821–71.
- Hassani Sadi M, Shahidi Bonjar GH. Plants used in folkloric medicine of Iran are exquisite bio-resources in production of silver nanoparticles. IET nanobiotechnology. 2017;11(3):300–309.
- Hassanisaadi M, Bonjar GHS, Rahdar A, Pandey S, Hosseinipour A, Abdolshahi R. Environmentally safe biosynthesis of gold nanoparticles using plant water extracts. Nanomaterials. 2021;11(8):2033.
- Roopan SM, Madhumitha G, Rahuman AA, Kamaraj C, Bharathi A, Surendra TV. Low-cost and eco-friendly phyto-synthesis of silver nanoparticles using *Cocos nucifera* coir extract and its larvicidal activity. Ind Crops Prod. 2013;43:631–5.
- Karthiga P. Preparation of silver nanoparticles by garcinia mangostana stem extract and investigation of the antimicrobial properties. Biotechnol Res Innov. 2018;2:30–6.
- Dwivedi AD, Gopal K. Biosynthesis of silver and gold nanoparticles using chenopodium album leaf extract. Colloids Surf A Physicochem Eng Asp. 2010;369:27–33.
- Ahmad H, Rajagopal K, Shah AH, Bhat AH, Venugopal K. Study of bio-fabrication of iron nanoparticles and their fungicidal property against phytopathogens of apple orchards. IET Nanobiotechnol. 2017;11(3):230–5.
- 34. Ahmad H, Venugopal K, Rajagopal K, De Britto S, Nandini B, Pushpalatha HG, et al. Green synthesis and characterization of zinc oxide nanoparticles using *Eucalyptus globules* and their fungicidal ability against pathogenic fungi of apple orchards. Biomolecules. 2020;10(3):425.
- 35. Ahmad H, Venugopal K, Bhat AH, Kavitha K, Ramanan A, Rajagopal K, et al. Enhanced biosynthesis synthesis of copper oxide nanoparticles (CuO-NPs) for their antifungal activity toxicity against major phytopathogens of apple orchards. Pharm Res. 2020;37:1–12.
- Ijaz M, Zafar M, Iqbal T. Green synthesis of silver nanoparticles by using various extracts: a review. Inorganic Nano-Metal Chem. 2020;51:744–55.
- Shiraz M, Imtiaz H, Azam A, Hayat S. Phytogenic nanoparticles: synthesis, characterization, and their roles in physiology and biochemistry of plants. Biometals. 2024;37(1):23–70.
- Sapsford KE, Tyner KM, Dair BJ, Deschamps JR, Medintz IL. Analyzing nanomaterial bioconjugates: a review of current and emerging purification and characterization techniques. Anal Chem. 2011;83(12):4453–88.
- Rodríguez-Félix F, Del-Toro-Sánchez CL, Tapia-Hernández JA. A new design for obtaining white zein micro- and nanoparticles powder: antisolvent-dialysis method. Food Sci Biotechnol. 2020;29:619–29.
- Hassani Sadi M, Shahidi Bonjar GH. Plants used in folkloric medicine of Iran are exquisite bio-resources in production of silver nanoparticles. IET Nanobiotechnol. 2017;11(3):300–9.
- Zee DZ, MacRenaris KW, O'Halloran TV. Quantitative imaging approaches to understanding biological processing of metal ions. Curr Opin Chem Biol. 2022;69: 102152.
- Allen C, Qiu TA, Pramanik S, Buchman JT, Krause MOP, Murphy CJ. Research highlights: investigating the role of nanoparticle surface charge in nano-bio interactions. Environ Sci Nano. 2017;4:741–6. https://doi.org/10.1039/C7EN90014G.
- Kumah EA, Fopa RD, Harati S, Boadu P, Zohoori FV, Pak T. Human and environmental impacts of nanoparticles: a scoping review of the current literature. BMC Public Health. 2023;23(1):1059.
- Sarkar A, Chakraborty N, Acharya K. Chitosan nanoparticles mitigate alternaria leaf spot disease of chilli in nitric oxide dependent way. Plant Physiol Biochem. 2022;180:64–73.
- Sarkar MR, Rashid MHO, Rahman A, Kafi MA, Hosen MI, Rahman MS, Khan MN. Recent advances in nanomaterials based sustainable agriculture: an overview. Environ Nanotechnol Monitor Manag. 2022;18: 100687.
- Tripathi S, Mahra S, Tiwari K, Rana S, Tripathi DK, Sharma S, Sahi S. Recent advances and perspectives of nanomaterials in agricultural management and associated environmental risk: a review. Nanomaterials. 2023;13(10):1604.
- Kashyap PL, Kumar S, Srivastava AK. Nanodiagnostics for plant pathogens. Environ Chem Lett. 2017;15:7–13.
- Chen Y, Kistler HC, Ma Z. Fusarium graminearum trichothecene mycotoxins: biosynthesis, regulation, and management. Annu Rev Phytopathol. 2019;57:15–39.

- 49. Bhattacharjee R, Kumar L, Mukerjee N, Anand U, Dhasmana A, Preetam S, Bhaumik S, Sihi S, Pal S, Khare T. The emergence of metal oxide nano-particles (NPs) as a phytomedicine: a two-facet role in plant growth, nano-toxicity and anti-phyto-microbial activity. Biomed Pharmacother. 2022;155: 113658.
- El-Sayed ESR, Mohamed SS, Mousa SA, El-Seoud MAA, Elmehlawy AA, Abdou DAM. Bifunctional role of some biogenic nanoparticles in controlling wilt disease and promoting growth of common bean. AMB Expr. 2023. https://doi.org/10.1186/S13568-023-01546-7.
- Malandrakis AA, Kavroulakis N, Chrysikopoulos CV. Use of copper, silver and zinc nanoparticles against foliar and soil-borne plant pathogens. Sci Total Environ. 2019;670:292–9.
- Bhargava P, Kumar A, Kumar S, Azad CS. Impact of fungicides and nanoparticles on ustilaginoidea virens causing false smut disease of rice. J Pharmacogn Phytochem. 2018;7:1541–4.
- Alotaibi MO, Alotaibi NM, Ghoneim AM, Ul Ain N, Irshad MA, Nawaz R, Abbas T, Abbas A, Rizwan M, Ali S. Effect of green synthesized cerium oxide nanoparticles on fungal disease of wheat plants: a field study. Chemosphere. 2023;339:139731.
- Shende S, Bhagat R, Raut R, Rai M, Gade A. Myco-fabrication of copper nanoparticles and its effect on crop pathogenic fungi. IEEE Trans Nanobiosci. 2021;20:146–53.
- Shah V, Dobiášová P, Baldrian P, Nerud F, Kumar A, Seal S. Influence of iron and copper nanoparticle powder on the production of lignocellulose degrading enzymes in the fungus trametes versicolor. J Hazard Mater. 2010;178:1141–5.
- Bramhanwade K, Shende S, Bonde S, Gade A, Rai M. Fungicidal activity of Cu nanoparticles against fusarium causing crop diseases. Environ Chem Lett. 2016;14:229–35.
- 57. Kailasa SK, Park T-J, Rohit JV, Koduru JR. Antimicrobial activity of silver nanoparticles. In: Nanoparticles in pharmacotherapy. Elsevier; 2019. p. 461–84.
- Bedlovičová Z, Strapáč I, Baláž M, Salayová A. A brief overview on antioxidant activity determination of silver nanoparticles. Molecules. 2020;25:3191.
- 59. Elmer W, White JC. The future of nanotechnology in plant pathology. Annu Rev Phytopathol. 2018;56:111–33.
- Khan MR, Rizvi TF, Ahamad F. Application of nanomaterials in plant disease diagnosis and management. Nanobiotechnol Appl Plant Protect. 2019;2:19–33.
- 61. Samy MA, Abbassy MA, Hafez EE, Rabea EI, Aseel DG. Biosynthesis and characterization of silver nanoparticles produced by plant extracts and its antimicrobial activity. South Asian J Res Microbiol. 2019;3:1–14.
- 62. Jo Y-K, Kim BH, Jung G. Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. Plant Dis. 2009;93:1037–43.
- Hassanisaadi M, Barani M, Rahdar A, Heidary M, Thysiadou A, Kyzas GZ. Role of agrochemical-based nanomaterials in plants: biotic and abiotic stress with germination improvement of seeds. Plant Growth Regul. 2022;97(2):375–418.
- Kalia A, Abd-Elsalam KA, Kuca K. Zinc-based nanomaterials for diagnosis and management of plant diseases: ecological safety and future prospects. J Fungi. 2020;6:222.
- 65. Helmy KG, Partila AM, Salah M. Gamma radiation and polyvinyl pyrrolidone mediated synthesis of zinc oxide/zinc sulfide nanoparticles and evaluation of their antifungal effect on pre and post harvested orange and pomegranate fruits. Biocatal Agric Biotechnol. 2020;29: 101728.
- 66. Panyuta O, Belava V, Fomaidi S, Kalinichenko O, Volkogon M, Taran N. The effect of pre-sowing seed treatment with metal nanoparticles on the formation of the defensive reaction of wheat seedlings infected with the eyespot causal agent. Nanoscale Res Lett. 2016;11:1–5.
- 67. Bouqellah NA, El-Sayyad GS, Attia MS. Induction of tomato plant biochemical immune responses by the synthesized zinc oxide nanoparticles against wilt-induced *Fusarium oxysporum*. Int Microbiol. 2023. https://doi.org/10.1007/S10123-023-00404-7.
- Riseh RS, Hassanisaadi M, Vatankhah M, Soroush F, Varma RS. Nano/ microencapsulation of plant biocontrol agents by chitosan, alginate, and other important biopolymers as a novel strategy for alleviating plant biotic stresses. Int J Biol Macromol. 2022;222:1589–604.
- 69. Zheng K, Lu J, Li J, Yu Y, Zhang J, He Z, Ismail OM, Wu J, Xie X, Li X, et al. Efficiency of chitosan application against phytophthora infestans and

the activation of defence mechanisms in potato. Int J Biol Macromol. 2021;182:1670–80. https://doi.org/10.1016/j.ijbiomac.2021.05.097.

- Hoang NH, le Thanh T, Sangpueak R, Treekoon J, Saengchan C, Thepbandit W, Papathoti NK, Kamkaew A, Buensanteai N. Chitosan nanoparticles-based ionic gelation method: a promising candidate for plant disease management. Polymers (Basel). 2022;14:662.
- Guerra-Sanchez MG, Vega-Pérez J, Velazquez-Del Valle MG, Hernandez-Lauzardo AN. Antifungal activity and release of compounds on *Rhizopus stolonifer* (Ehrenb.: Fr.) Vuill. by effect of chitosan with different molecular weights. Pestic Biochem Physiol. 2009;93:18–22.
- 72. Al-Mokadem AZ, Alnaggar AEAM, Mancy AG, Sofy AR, Sofy MR, Mohamed AKSH, Abou Ghazala MMA, El-Zabalawy KM, Salem NFG, Elnosary ME. Foliar application of chitosan and phosphorus alleviate the potato virus Y-induced resistance by modulation of the reactive oxygen species, antioxidant defense system activity and gene expression in potato. Agronomy. 2022;12:3064.
- Riseh RS, Hassanisaadi M, Vatankhah M, Babaki SA, Barka EA. Chitosan as potential natural compound to manage plant diseases. Int J Biol Macromol. 2022;220:998–1009.
- Muthukrishnan S, Murugan I, Selvaraj M. Chitosan nanoparticles loaded with thiamine stimulate growth and enhances protection against wilt disease in chickpea. Carbohydr Polym. 2019;212:169–77.
- Choudhary RC, Kumaraswamy RV, Kumari S, Sharma SS, Pal A, Raliya R, Biswas P, Saharan V. Cu-chitosan nanoparticle boost defense responses and plant growth in maize (*Zea mays* L.). Sci Rep. 2017;7:1–11.
- Vanti GL, Nargund VB, Vanarchi R, Kurjogi M, Mulla SI, Tubaki S, Patil RR. Synthesis of gossypium hirsutum-derived silver nanoparticles and their antibacterial efficacy against plant pathogens. Appl Organomet Chem. 2019;33: e4630.
- 77. Qasim M, Akhtar W, Haseeb M, Sajjad H, Rasheed M. Potential role of nanoparticles in plants protection. Life Sci J. 2022;19:31–8.
- Khan MR, Rizvi TF. Application of nanofertilizer and nanopesticides for improvements in crop production and protection. In: Nanoscience and plant–soil systems. Springer; 2017. p. 405–27.
- Kasprowicz MJ, Kozioł M, Gorczyca A. The effect of silver nanoparticles on phytopathogenic spores of *Fusarium culmorum*. Can J Microbiol. 2010;56:247–53.
- Javed B, Nadhman A. Optimization, characterization and antimicrobial activity of silver nanoparticles against plant bacterial pathogens phytosynthesized by *Mentha longifolia*. Mater Res ess. 2020;7: 085406.
- Kaur P, Thakur R, Duhan JS, Chaudhury A. Management of wilt disease of chickpea in vivo by silver nanoparticles biosynthesized by rhizospheric microflora of chickpea (*Cicer arietinum*). J Chem Technol Biotechnol. 2018;93:3233–43.
- 82. Elbeshehy EKF, Elazzazy AM, Aggelis G. Silver nanoparticles synthesis mediated by new isolates of *Bacillus* Spp., nanoparticle characterization and their activity against bean yellow mosaic virus and human pathogens. Front Microbiol. 2015;6:453.
- Murali M, Naziya B, Singh SB, Chandrashekar S, Udayashankar AC, Amruthesh KN. Management of plant fungal disease by microbial nanotechnology. In: Microbial nanotechnology: green synthesis and applications. Springer; 2021. p. 287–305.
- Elamawi RM, Al-Harbi RE, Hendi AA. Biosynthesis and characterization of silver nanoparticles using trichoderma longibrachiatum and their effect on phytopathogenic fungi. Egypt J Biol Pest Control. 2018;28:1–11.
- Kora AJ, Mounika J, Jagadeeshwar R. Rice leaf extract synthesized silver nanoparticles: an in vitro fungicidal evaluation against rhizoctonia solani, the causative agent of sheath blight disease in rice. Fungal Biol. 2020;124:671–81.
- Salem SS, Ali OM, Reyad AM, Abd-Elsalam KA, Hashem AH. Pseudomonas indica-mediated silver nanoparticles: antifungal and antioxidant biogenic tool for suppressing mucormycosis fungi. J Fungi. 2022;8:126.
- Gabal E, Ramadan MM, Alghuthaymi MA, Abd-Elsalam KA. Copper Nanostructures applications in plant protection. In: Nanobiotechnology applications in plant protection. Springer; 2018. p. 63–86.
- Bhuvaneshwari V, Ramasamy NK, Kumar SI, Kalaivani S, Vaidehi D, Kumar DK. Antimicrobial activity of copper nanomaterials: current status and future perspectives. In: Copper nanostructures: next-generation of agrochemicals for sustainable agroecosystems. Elsevier; 2022. p. 453–75.

- Oussou-Azo AF, Nakama T, Nakamura M, Futagami T, Vestergaard MCM. Antifungal potential of nanostructured crystalline copper and its oxide forms. Nanomaterials. 2020;10:1003.
- Ahmed T, Wu Z, Jiang H, Luo J, Noman M, Shahid M, Manzoor I, Allemailem KS, Alrumaihi F, Li B. Bioinspired green synthesis of zinc oxide nanoparticles from a native *bacillus cereus* Strain RNT6: characterization and antibacterial activity against rice panicle blight pathogens burkholderia glumae and B. Gladioli Nanomaterials. 2021;11:884.
- Kumar A, Choudhary A, Kaur H, Guha S, Mehta S, Husen A. Potential applications of engineered nanoparticles in plant disease management: a critical update. Chemosphere. 2022;295: 133798. https://doi. org/10.1016/j.chemosphere.2022.133798.
- Hossain A, Abdallah Y, Ali MA, Masum MMI, Li B, Sun G, Meng Y, Wang Y, An Q. Lemon-fruit-based green synthesis of zinc oxide nanoparticles and titanium dioxide nanoparticles against soft rot bacterial pathogen *Dickeya dadantii*. Biomolecules. 2019;9:863.
- Satti ⁵H, Raja NI, Javed B, Akram A, Mashwani Z-R, Ahmad MS, Ikram M. Titanium dioxide nanoparticles elicited agro-morphological and physicochemical modifications in wheat plants to control Bipolaris Sorokiniana. PLoS ONE. 2021;16: e0246880.
- 94. Singh R, Kuddus M, Singh PK, Choden D. Nanotechnology for nanophytopathogens: from detection to the management of plant viruses. Biomed Res Int 2022;2022:12. Article ID 8688584. https://doi.org/10. 1155/2022/8688584.
- Oh J-W, Chun SC, Chandrasekaran M. Preparation and in vitro characterization of chitosan nanoparticles and their broad-spectrum antifungal action compared to antibacterial activities against phytopathogens of tomato. Agronomy. 2019;9:21.
- Ahmed T, Noman M, Luo J, Muhammad S, Shahid M, Ali MA, Zhang M, Li B. Bioengineered chitosan-magnesium nanocomposite: a novel agricultural antimicrobial agent against *Acidovorax oryzae* and *Rhizoctonia solani* for sustainable rice production. Int J Biol Macromol. 2021;168:834–45.
- Lata C, Kumar N, Kaur G, Rani R, Pundir P, Rana AS. Applications of nanobiotechnological approaches in diagnosis and protection of wheat diseases. In: Cereal diseases: nanobiotechnological approaches for diagnosis and management. Springer; 2022. p. 345–70.
- Irshad MA, Nawaz R, Ur Rehman MZ, Imran M, Ahmad J, Ahmad S, Inam A, Razzaq A, Rizwan M, Ali S. Synthesis and characterization of titanium dioxide nanoparticles by chemical and green methods and their antifungal activities against wheat rust. Chemosphere. 2020;258:127352.
- Omar HS, Al Mutery A, Osman NH, Reyad NEHA, Abou-Zeid MA. Genetic diversity, antifungal evaluation and molecular docking studies of cuchitosan nanoparticles as prospective stem rust inhibitor candidates among some Egyptian wheat genotypes. PLoS ONE. 2021;16:e0257959.
- Pham TT, Nguyen TH, Thi TV, Nguyen T-T, Le TD, Vo DMH, Nguyen DH, Nguyen CK, Nguyen DC, Nguyen TT. Investigation of chitosan nanoparticles loaded with protocatechuic acid (PCA) for the resistance of *Pyricularia oryzae* fungus against rice blast. Polymers (Basel). 2019;11:177.
- 101. Abdallah Y, Ogunyemi SO, Abdelazez A, Zhang M, Hong X, Ibrahim E, Hossain A, Fouad H, Li B, Chen J. The green synthesis of MgO nanoflowers using *Rosmarinus officinalis* L. (Rosemary) and the antibacterial activities against *Xanthomonas oryzae* Pv. Oryzae. Biomed Res Int. 2019;2019:5620989.
- 102. Cui J, Liang Y, Yang D, Liu Y. Facile fabrication of rice husk based silicon dioxide nanospheres loaded with silver nanoparticles as a rice antibacterial agent. Sci Rep. 2016;6:1–10.
- Shenashen M, Derbalah A, Hamza A, Mohamed A, el Safty S. Recent trend in controlling root rot disease of tomato caused by *Fusarium solani* using aluminasilica nanoparticles. Int J Adv Res Biol Sci. 2017;4:105–19.
- Sathiyabama M, Charles RE. Fungal cell wall polymer based nanoparticles in protection of tomato plants from wilt disease caused by *Fusarium oxysporum* f. Sp. Lycopersici. Carbohydr Polym. 2015;133:400–7.
- Joshi SM, de Britto S, Jogaiah S. Myco-engineered selenium nanoparticles elicit resistance against tomato late blight disease by regulating differential expression of cellular, biochemical and defense responsive genes. J Biotechnol. 2021;325:196–206.
- 106. van Loon LC. The intelligent behavior of plants. Trends Plant Sci. 2016;21:286–94.

- Rajwade JM, Chikte RG, Paknikar KM. Nanomaterials: new weapons in a crusade against phytopathogens. Appl Microbiol Biotechnol. 2020;104:1437–61.
- Abdelrhim AS, Mazrou YSA, Nehela Y, Atallah OO, El-Ashmony RM, Dawood MFA. Silicon dioxide nanoparticles induce innate immune responses and activate antioxidant machinery in wheat against *Rhizoctonia solani*. Plants. 2021;10:2758.
- 109. Warghane A, Saini R, Shri M, Andankar I, Ghosh DK, Chopade BA. Application of nanoparticles for management of plant viral pathogen: current status and future prospects. Virology. 2024;592:109998.
- 110. Cai L, Cai L, Jia H, Liu C, Wang D, Sun X. Foliar exposure of Fe3O4 nanoparticles on *Nicotiana benthamiana*: evidence for nanoparticles uptake, plant growth promoter and defense response elicitor against plant virus. J Hazard Mater. 2020;393: 122415. https://doi.org/10.1016/j.jhazm at.2020.122415.
- 111. Liu Q, Li Y, Xu K, Li D, Hu H, Zhou F, et al. Clay nanosheet-mediated delivery of recombinant plasmids expressing artificial miRNAs via leaf spray to prevent infection by plant DNA viruses. Horticult Res. 2020;7:179.
- 112. Mitter N, et al. Clay nanosheets for topical delivery of RNAi for sustained protection against plant viruses. Nat Plants. 2017;3:16207.
- 113. Anjum S, Anjum I, Hano C, Kousar S. Advances in nanomaterials as novel elicitors of pharmacologically active plant specialized metabolites: current status and future outlooks. RSC Adv. 2019;9:40404–23.
- 114. Imada K, Sakai S, Kajihara H, Tanaka S, Ito S. Magnesium oxide nanoparticles induce systemic resistance in tomato against bacterial wilt disease. Plant Pathol. 2016;65:551–60.
- 115. Paul A, Roychoudhury A. Go green to protect plants: repurposing the antimicrobial activity of biosynthesized silver nanoparticles to combat phytopathogens. Nanotechnol Environ Eng. 2021;6:1–22.
- Saharan V, Mehrotra A, Khatik R, Rawal P, Sharma SS, Pal A. Synthesis of chitosan based nanoparticles and their in vitro evaluation against phytopathogenic fungi. Int J Biol Macromol. 2013;62:677–83.
- El-Shetehy M, Wang C, Shine MB, Yu K, Kachroo A, Kachroo P. Nitric oxide and reactive oxygen species are required for systemic acquired resistance in plants. Plant Signal Behav. 2015;10: e998544.
- Cao X, Wang C, Luo X, Yue L, White JC, Elmer W, Dhankher OP, Wang Z, Xing B. Elemental sulfur nanoparticles enhance disease resistance in tomatoes. ACS Nano. 2021;15(7):11817–27.
- Kandhol N, Singh VP, Peralta-Videa J, Corpas FJ, Tripathi DK. Silica nanoparticles: the rising star in plant disease protection. Trends Plant Sci. 2022;27:7–9. https://doi.org/10.1016/j.tplants.2021.10.007.
- 120. Luo X, Cao X, Wang C, Yue L, Chen X, Yang H, Le X, Zhao X, Wu F, Wang Z. Nitrogen-doped carbon dots alleviate the damage from tomato bacterial wilt syndrome: systemic acquired resistance activation and reactive oxygen species scavenging. Environ Sci Nano. 2021;8:3806–19.
- 121. Li Z, Juneau P, Lian Y, Zhang W, Wang S, Wang C, Shu L, Yan Q, He Z, Xu K. Effects of titanium dioxide nanoparticles on photosynthetic and antioxidative processes of *Scenedesmus obliquus*. Plants. 2020;9:1748.
- 122. Martin RE, Postiglione AE, Muday GK. Reactive oxygen species function as signaling molecules in controlling plant development and hormonal responses. Curr Opin Plant Biol. 2022;69: 102293.
- 123. Marslin G, Sheeba CJ, Franklin G. Nanoparticles alter secondary metabolism in plants via ROS burst. Front Plant Sci. 2017;8:832.
- 124. Barbaś P, Skiba D, Pszczółkowski P, Sawicka B. Mechanisms of plant natural immunity and the role of selected oxylipins as molecular mediators in plant protection. Agronomy. 2022;12:2619.
- 125. de Pereira AES, Oliveira HC, Fraceto LF. Polymeric nanoparticles as an alternative for application of gibberellic acid in sustainable agriculture: a field study. Sci Rep. 2019;9:1–10.
- Du Z, Li M, Zou F, Song Y, Xu S, Wu L, Li L, Li G. VO2@ SiO2 nanoparticlebased films with localized surface plasmon resonance for smart windows. ACS Appl Nano Mater. 2022;5:12972–9.
- Clemente I, Menicucci F, Colzi I, Sbraci L, Benelli C, Giordano C, Gonnelli C, Ristori S, Petruccelli R. Unconventional and sustainable nanovectors for phytohormone delivery: insights on *Olea europaea*. ACS Sustain Chem Eng. 2018;6:15022–31.
- 128. Siddaiah CN, Prasanth KVH, Satyanarayana NR, Mudili V, Gupta VK, Kalagatur NK, Satyavati T, Dai X-F, Chen J-Y, Mocan A. Chitosan nanoparticles having higher degree of acetylation induce resistance against pearl millet downy mildew through nitric oxide generation. Sci Rep. 2018;8:1–14.

- Mondaca F, Mtz-Enriquez AI, Pariona N. Copper-based nanostructures for plant disease management. In: Copper nanostructures: next-generation of agrochemicals for sustainable agroecosystems. Elsevier; 2022. p. 185–201.
- 130. Vorster BJ, Cullis CA, Kunert KJ. Plant vacuolar processing enzymes. Front Plant Sci. 2019;10:479.
- 131. Chaudhary P, Khati P, Gangola S, Kumar A, Kumar R, Sharma A. Impact of nanochitosan and *Bacillus* Spp. on health, productivity and defence response in *Zea mays* under field condition. 3 Biotech. 2021;11:1–11.
- 132. Kumaraswamy RV, Kumari S, Choudhary RC, Pal A, Raliya R, Biswas P, Saharan V. Engineered chitosan based nanomaterials: bioactivities, mechanisms and perspectives in plant protection and growth. Int J Biol Macromol. 2018;113:494–506.
- 133. Oliveira MDM, Varanda CMR, Félix MRF. Induced resistance during the interaction pathogen x plant and the use of resistance inducers. Phytochem Lett. 2016;15:152–8.
- Narasimhamurthy K, Udayashankar AC, de Britto S, Lavanya SN, Abdelrahman M, Soumya K, Shetty HS, Srinivas C, Jogaiah S. Chitosan and chitosan-derived nanoparticles modulate enhanced immune response in tomato against bacterial wilt disease. Int J Biol Macromol. 2022;220:223–37.
- Sathiyabama M, Indhumathi M. Chitosan thiamine nanoparticles intervene innate immunomodulation during chickpea-Fusarium interaction. Int J Biol Macromol. 2022;198:11–7.
- Liang W, Yu A, Wang G, Zheng F, Hu P, Jia J, Xu H. A novel water-based chitosan-la pesticide nanocarrier enhancing defense responses in rice (*Oryza sativa* L.) growth. Carbohydr Polym. 2018;199:437–44. https://doi. org/10.1016/j.carbpol.2018.07.042.
- Cai L, Liu C, Fan G, Liu C, Sun X. Preventing viral disease by ZnONPs through directly deactivating TMV and activating plant immunity in *Nicotiana benthamiana*. Environ Sci Nano. 2019;6:3653–69.
- Derbalah ASH, Elsharkawy MM. A new strategy to control cucumber mosaic virus using fabricated NiO-nanostructures. J Biotechnol. 2019;306:134–41.
- Noha K, Bondok AM, El-Dougdoug KA. Evaluation of silver nanoparticles as antiviral agent against ToMV and PVY in tomato plants. Sciences. 2018;8:100–11.
- 140. Zhang H, Du W, Peralta-Videa JR, Gardea-Torresdey JL, White JC, Keller A, Guo H, Ji R, Zhao L. Metabolomics reveals how cucumber (*Cucumis sativus*) reprograms metabolites to cope with silver ions and silver nanoparticle-induced oxidative stress. Environ Sci Technol. 2018;52:8016–26.
- Sarkar J, Chakraborty N, Chatterjee A, Bhattacharjee A, Dasgupta D, Acharya K. Green synthesized copper oxide nanoparticles ameliorate defence and antioxidant enzymes in lens culinaris. Nanomaterials. 2020;10:312.
- 142. Sofy AR, Sofy MR, Hmed AA, Dawoud RA, Alnaggar AE-AM, Soliman AM, El-Dougdoug NK. Ameliorating the adverse effects of tomato mosaic tobamovirus infecting tomato plants in egypt by boosting immunity in tomato plants using zinc oxide nanoparticles. Molecules. 2021;26:1337.
- Méndez-López A, González-García Y, Juárez-Maldonado A. Stimulatory role of nanomaterials on agricultural crops. In: Nano-enabled Agrochemicals in agriculture. Elsevier; 2022. p. 219–46.
- 144. Ch G, Ntalli N, Menkissoglu-Spiroudi U, Dendrinou-Samara C. Essential metal-based nanoparticles (copper/iron NPs) as potent nematicidal agents against *Meloidogyne* Spp. J Nanotechnol Res. 2019;1:44–58.
- 145. Khater M, de La Escosura-Muñiz A, Merkoçi A. Biosensors for plant pathogen detection. Biosens Bioelectron. 2017;93:72–86.
- Kuska MT, Heim RHJ, Geedicke I, Gold KM, Brugger A, Paulus S. Digital plant pathology: a foundation and guide to modern agriculture. J Plant Dis Protect. 2022;129:1–12.
- 147. Kulabhusan PK, Tripathi A, Kant K. Gold nanoparticles and plant pathogens: an overview and prospective for biosensing in forestry. Sensors. 2022;22:1259.
- Neelam A, Tabassum S. Optical sensing technologies to elucidate the interplay between plant and microbes. Micromachines. 2023;14(1):195.
- Jez JM, Topp CN, Silva G, Tomlinson J, Onkokesung N, Sommer S, Mrisho L, Legg J, Adams IP, Gutierrez-Vazquez Y. Plant pest surveillance: from satellites to molecules. Emerg Top Life Sci. 2021;5:275–87.
- 150. Cardoso RM, Pereira TS, Facure MHM, dos Santos DM, Mercante LA, Mattoso LHC, Correa DS. Current progress in plant pathogen detection

enabled by nanomaterials-based (bio) sensors. Sens Actuators Rep. 2021;4:100068.

- Naresh V, Lee N. A review on biosensors and recent development of nanostructured materials-enabled biosensors. Sensors. 2021;21:1109.
- 152. Manjunatha RL, Naik D, Usharani KV. Nanotechnology application in agriculture: a review. J Pharmacogn Phytochem. 2019;8:1073–83.
- 153. Singh S, Singh M, Agrawal VV, Kumar A. An attempt to develop surface plasmon resonance based immunosensor for Karnal Bunt (Tilletia Indica) diagnosis based on the experience of nano-gold based lateral flow immuno-dipstick test. Thin Solid Films. 2010;519:1156–9.
- Kashyap PL, Kumar S, Jasrotia P, Singh DP, Singh GP. Nanosensors for plant disease diagnosis: current understanding and future perspectives. In: Nanoscience for sustainable agriculture. Springer; 2019. p. 189–205.
- 155. Mathivanan, S. Perspectives of Nano-materials and nanobiosensors in food safety and agriculture. Novel nanomaterials. 2021. p. 197.
- 156. Feng H, Gonzalez Viejo C, Vaghefi N, Taylor PWJ, Tongson E, Fuentes S. Early detection of *Fusarium oxysporum* infection in processing tomatoes (*Solanum lycopersicum*) and pathogen–soil interactions using a lowcost portable electronic nose and machine learning modeling. Sensors. 2022;22:8645.
- 157. Jiarpinijnun A, Osako K, Siripatrawan U. Visualization of volatomic profiles for early detection of fungal infection on storage jasmine brown rice using electronic nose coupled with chemometrics. Measurement. 2020;157: 107561.
- 158. Oerke E-C. Remote sensing of diseases. Annu Rev Phytopathol. 2020;58:225–52.
- Che'Ya NN, Mohidem NA, Roslin NA, Saberioon M, Tarmidi MZ, Arif Shah J, Fazlil Ilahi WF, Man N. Mobile computing for pest and disease management using spectral signature analysis: a review. Agronomy. 2022;12:967.
- Adama A, Ee KP, Sahari N, Tida A, Shang CY, Tawie KM, Kamarudin S, Mohamad H. LADA: diagnosing black pepper pest and diseases with decision tree. Int J Adv Sci Eng Inf Technol. 2018;8:1584–90.
- Liao Y-C, Pingli H, Senzhao C, Yao M-J, Zhang J-B, Liu J-L. Plantibodies: a novel strategy to create pathogen-resistant plants. Biotechnol Genet Eng Rev. 2006;23:253–72. https://doi.org/10.1080/02648725.2006.10648 087.
- 162. Ma JK, Drossard J, Lewis D, Altmann F, Boyle J, Christou P, Cole T, Dale P, van Dolleweerd CJ, Isitt V, et al. Regulatory approval and a first-inhuman phase I clinical trial of a monoclonal antibody produced in transgenic tobacco plants. Plant Biotechnol J. 2015;13:1106–20. https:// doi.org/10.1111/pbi.12416.
- 163. De Jaeger G, De Wilde C, Eeckhout D, Fiers E, Depicker A. The plantibody approach: expression of antibody genes in plants to modulate plant metabolism or to obtain pathogen resistance. Plant Mol Biol. 2000;43:419–28. https://doi.org/10.1023/A:1006471528756.
- Sanzari I, Leone A, Ambrosone A. Nanotechnology in plant science: to make a long story short. Front Bioeng Biotechnol. 2019;7:120.
- Pariona N, Mtz-Enriquez AI, Sánchez-Rangel D, Carrión G, Paraguay-Delgado F, Rosas-Saito G. Green-synthesized copper nanoparticles as a potential antifungal against plant pathogens. RSC Adv. 2019;9:18835–43.
- 166. Sembada AA, Lenggoro IW. Transport of nanoparticles into plants and their detection methods. Nanomaterials. 2024;14(2):131.
- 167. Wang X, Xie H, Wang P, Yin H. Nanoparticles in plants: uptake, transport and physiological activity in leaf and root. Materials. 2023;16(8):3097.
- Kumar V, Sharma M, Khare T, Wani SH. Impact of nanoparticles on oxidative stress and responsive antioxidative defense in plants. In: Nanomaterials in plants, algae, and microorganisms. Academic Press; 2018. p. 393–406.
- Akhtar N, Khan S, Rehman SU, Rehman ZU, Khatoon A, Rha ES, Jamil M. Synergistic effects of zinc oxide nanoparticles and bacteria reduce heavy metals toxicity in rice (*Oryza sativa* L.) plant. Toxics. 2021;9(5):113.
- 170. McKee MS, Filser J. Impacts of metal-based engineered nanomaterials on soil communities. Environ Sci Nano. 2016;3(3):506–33.
- 171. Grun AL, Straskraba S, Schulz S, Schloter M, Emmerling C. Longterm effects of environmentally relevant concentrations of silver nanoparticles on microbial biomass, enzyme activity, and functional genes involved in the nitrogen cycle of loamy soil. J Environ Sci. 2018;69:12–22.

- 172. Khan ST, Adil SF, Shaik MR, Alkhathlan HZ, Khan M, Khan M. Engineered nanomaterials in soil: their impact on soil microbiome and plant health. Plants. 2021;11(1):109.
- 173. Hao Y, Ma C, Zhang Z, Song Y, Cao W, Guo J, et al. Carbon nanomaterials alter plant physiology and soil bacterial community composition in a rice-soil-bacterial ecosystem. Environ Pollut. 2018;232:123–36.
- 174. An C, Sun C, Li N, Huang B, Jiang J, Shen Y, Wang C, Zhao X, Cui B, Wang C. Nanomaterials and nanotechnology for the delivery of agrochemicals: strategies towards sustainable agriculture. J Nanobiotechnology. 2022;20:1–19.
- 175. Lin W, Hu X, Zhang W, Rogers WJ, Cai W. Hydrogen Peroxide mediates defence responses induced by chitosans of different molecular weights in rice. J Plant Physiol. 2005;162:937–44.
- 176. Singh RP, Handa R, Manchanda G. Nanoparticles in sustainable agriculture: an emerging opportunity. J Control Release. 2021;329:1234–48. https://doi.org/10.1016/j.jconrel.2020.10.051.
- Grillo R, Fraceto LF, Amorim MJB, Scott-Fordsmand JJ, Schoonjans R, Chaudhry Q. Ecotoxicological and regulatory aspects of environmental sustainability of nanopesticides. J Hazard Mater. 2021;404: 124148. https://doi.org/10.1016/j.jhazmat.2020.124148.
- Konappa N, Krishnamurthy S, Arakere UC, Chowdappa S, Akbarbasha R, Ramachandrappa NS. Nanofertilizers and nanopesticides: recent trends, future prospects in agriculture. In: Advances in nano-fertilizers and nano-pesticides in agriculture. Elsevier; 2021. p. 281–330.
- Singh D, Gurjar BR. Nanotechnology for agricultural applications: facts, issues, knowledge gaps, and challenges in environmental risk assessment. J Environ Manage. 2022;322: 116033. https://doi.org/10.1016/j. jenvman.2022.116033.
- Shelar A, Nile SH, Singh AV, Rothenstein D, Bill J, Xiao J, et al. Recent advances in nano-enabled seed treatment strategies for sustainable agriculture: challenges, risk assessment, and future perspectives. Nano-Micro Letters. 2023;15(1):54.
- Onyeaka H, Passaretti P, Miri T, Al-Sharify ZT. The safety of nanomaterials in food production and packaging. Curr Res Food Sci. 2022;5:763–74.
- Okeke ES, Ezeorba TPC, Mao G, Chen Y, Feng W, Wu X. Nano-enabled agrochemicals/materials: potential human health impact, risk assessment, management strategies and future prospects. Environ Pollut. 2022;295: 118722.
- Yan A, Chen Z. Impacts of silver nanoparticles on plants: a focus on the phytotoxicity and underlying mechanism. Int J Mol Sci. 2019;20:1003.
- Tripathi DK, Singh S, Singh S, Pandey R, Singh VP, Sharma NC, Prasad SM, Dubey NK, Chauhan DK. An overview on manufactured nanoparticles in plants: uptake, translocation, accumulation and phytotoxicity. Plant Physiol Biochem. 2017;110:2–12.
- Avila-Quezada GD, Golinska P, Rai M. Engineered nanomaterials in plant diseases: can we combat phytopathogens? Appl Microbiol Biotechnol. 2022;106(1):117–29.
- Mittal D, Kaur G, Singh P, Yadav K, Ali SA. Nanoparticle-based sustainable agriculture and food science: recent advances and future outlook. Front Nanotechnol. 2020;2: 579954.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.